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(54) NOVEL HEMOPOIETIN RECEPTOR PROTEINS

(57) The present invention provides novel hemopoietin receptor proteins (proteins comprising the amino acid sequence of SEQ ID NOs: 1, 3, 5, 7, 19, or 21), proteins comprising a modified amino acid sequence of the amino acid sequence of the above protein in which one or more amino acids have been deleted, added, and/or replaced with another amino acid, genes encoding these proteins, methods of producing the proteins, as well as uses of these proteins and genes.

Descripti n

Technical Field

[0001] The present invention relates to novel hemopoietin receptor proteins, the encoding genes, and methods of production and uses thereof.

Background Art

[0002] A large number of cytokines are known as humoral factors that are involved in the proliferation/differentiation of various cells, or activation of differentiated mature cells, and also cell death. These cytokines have their own specific receptors, which are categorized into several families based on their structural similarities (Hilton D.J., in "Guidebook to Cytokines and Their Receptors" edited by Nicola N.A. (A Sambrook & Tooze Publication at Oxford University Press), 1994, p8-16).

[0003] Compared to similarities between receptors, primary-structure homology is quite low between cytokines, and a significant amino acid homology cannot be seen even among cytokine members that belong to the same receptor family. This explains the functional specificity of each cytokine, as well as similarities of cellular reactions induced by each cytokine.

[0004] Representative examples of the above-mentioned receptor families are the tyrosine kinase receptor family, hemopoletin receptor family, tumor necrosis factor (TNF) receptor family, and transforming growth factor β (TGF β) receptor family. Different signal transduction pathways have been reported to be involved in each of these families. Among these receptor families, many receptors of especially the hemopoletin receptor family are expressed in blood cells and immunocytes, and their ligands, cytokines, are often termed as hemopoletic factors or interleukins. Some of these hemopoletic factors or interleukins exist within blood and are thought to be involved in a systemic humoral regulation of hemopoletic or immune functions.

[0005] This contrasts with the belief that cytokines belonging to other families are often involved in only topical regulations. Some of these hemopoietins can be taken as hormone-like factors, and conversely, representative peptide hormones such as the growth hormone, prolactin, or leptin receptors also belong to the hemopoietin receptor family. Because of these hormone-like systemic regulatory features, it is anticipated that hemopoietin administration would be applied in the treatment of various diseases.

[0006] Among the large number of cytokines, those that are actually being clinically applied are, erythropoietin, G-CSF, GM-CSF, and IL-2. Combined with IL-11, LIF, and IL-12 that are being considered for clinical trials, and the above-mentioned peptide hormones such as growth hormone and prolactin, it can be envisaged that by searching among the above-mentioned various receptor families for a novel cytokine that binds to hemopoietin receptors, it is possible to find a cytokine that can be clinically applied with a higher efficiency.

[0007] As mentioned above, cytokine receptors have structural similarities between the family members. Using these similarities, many investigations are being carried out aiming at finding novel receptors. Regarding the tyrosine kinase receptor especially, many receptors have already been cloned using its highly conserved sequence at the catalytic site (Matthews W. et al., Cell, 1991, 65 (7) p1143-52). Compared to this, hemopoietin receptors do not have a tyrosine kinase-like enzyme activity domain in their cytoplasmic regions, and their signal transductions are known to be mediated through associations with other tyrosine kinase proteins existing freely in the cytoplasm.

[0008] Though the binding site on receptors associating with these cytoplasmic tyrosine kinases (JAK kinases) is conserved between family members, the homology is not very high (Murakami M. et al., Proc. Natl. Acad. Sci. USA, 1991, 88, 11349-11353). On one hand, the sequence that characterizes these hemopoietin receptors most well exists in the extracellular region, and especially the five amino acid Trp-Ser-Xaa-Trp-Ser (where Xaa is an arbitrary amino acid) motif is conserved in almost all of the hemopoietin receptors. Therefore, novel receptors are expected to be obtained by searching novel family members using this sequence. In fact, this approach has already identified the IL-11 receptor (Robb, L. et al., J. Biol. Chem., 1996, 271 (23) 13754-13761), leptin receptor (Gainsford T. et al., Proc. Natl. Acad. Sci. USA, 1996, 93 (25) p14564-8) and the IL-13 receptor (Hilton D.J. et al., Proc. Natl. Acad. Sci. USA, 1996, 93 (1) p497-501).

Disclosure of the Invention

[0009] The present invention provides a novel hemopoietin receptor protein, and the encoding DNA. The present invention also provides, a vector into which the DNA has been inserted, a transformant harboring the DNA, and a method of producing a recombinant protein using the transformant. It also provides a method of screening a compound that binds to the protein.

[0010] Until now, the inventors have been trying to search for a novel receptor using an oligonucleotide encoding

the Trp-Ser-Xaa-Trp-Ser motif as a probe by plaque hybridization, RT-PCR method, and so on. However, because of reasons such as the oligonucleotide tggag (t/c) nnntggag (t/c) (where n is an arbitrary nucleotide) that encodes the motif being short having just 15 nucleotides, and the g/c being high, it was extremely difficult to strictly select only those in which the 15 nucleotides have completely hybridized under the usual hybridization conditions.

[0011] Also, a similar sequence is contained within cDNA encoding proteins other than hemopoietin receptors, starting with various collagens that are thought to be widely distributed and also have high expression amounts, which makes the screening by the above-mentioned plaque hybridization and RT-PCR highly inefficient.

[0012] To solve these problems, and to estimate how many different hemopoietic receptor genes actually exist on the human genome, the inventors computer-searched sequences that completely coincided with each probe using all capable oligonucleotide sequences encoding the above-mentioned Trp-Ser-Xaa-Trp-Ser motif as probes.

[0013] Next, among the clones identified by the above search, the nucleotide sequence around the probe sequence of human genome-derived clones (cosmid, BAC, PAC) was converted to the amino acid sequence and compared with the amino acid sequence of known hemopoietin receptors to select human genes thought to encode hemopoietin receptor family members.

[0014] From the above search, two clones thought to be hemopoietin receptor genes were identified. One of these was the known GM-CSFβ receptor gene (derived from the 22q12.3-13.2 region of chromosome no. 22), and the other (BAC clone AC002303 derived from the 16p12 region of chromosome no. 16) was presumed to encode a novel hemopoietin receptor protein, and this human gene was named "NR8."

[0015] Next, the cDNA thought to encode NR8 was found within the human fetal liver cell cDNA library by RT-PCR using a specific primer designed based on the obtained nucleotide sequence. Furthermore, using this cDNA library as the template, the full-length cDNA NR8 α encoding a transmembrane receptor comprising 361 amino acids was ultimately obtained by 5'-RACE method and 3'-RACE method.

[0016] In the primary structure of NR8 α , a cysteine residue and a proline rich motif conserved between other family members, were well conserved in the extracellular region, and in the intracellular region, the Box 1 motif thought to be involved in signal transduction was well conserved, and therefore, NR8 α was thought to be a typical hemopoietin receptor.

[0017] Furthermore, the inventors revealed the presence of two genes named NR8 β and NR8 γ as selective splicing products of NR8 α .

[0018] The inventors next attempted the isolation of the mouse gene corresponding to NR8 gene. First, using an oligonucleotide primer designed within human NR8 cDNA sequence and a mouse brain cDNA library as the template, xenogeneic cross PCR cloning was done to isolate the mouse partial nucleotide sequence of the above receptor. Furthermore, based on the obtained partial sequence, an oligonucleotide primer was designed, and using this, the inventors succeeded in isolating the full-length ORF of the mouse homologous gene corresponding to NR8 by the 5'-RACE method and 3'-RACE method. As a result of determining the whole nucleotide sequence of the obtained cDNA clone, alike NR8, the presence of mouse NR8γ encoding a transmembrane receptor protein comprising 538 amino acids, and mouse NR8β encoding a secretory, soluble receptor-like protein comprising 144 amino acids were confirmed by the difference of transcripts derived from the splice variant. When the amino acid sequences encoded by these receptor genes were compared between human and mouse, a high homology of 98.9% was observed for NR8γ, and on the other hand, a homology of 97.2% was seen for NR8β as well. Furthermore, the inventors succeeded in isolating the objective positive clones by plaque screening against a mouse genomic DNA library using the obtained mouse NR8β cDNA fragment as the nmbe

[0019] Therefore, the present invention provides:

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- (1) a protein comprising the amino acid sequence from the 1st amino acid Met to the 361st amino acid Ser of SEQ ID NO: 1, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added, and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 361st amino acid Ser of SEQ ID NO: 1;
- (2) a protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 3, or a protein comprising a modified amino acid sequence of sald amino acid sequence in which one or more amino acids have been deleted, added, and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 3:
- (3) a protein comprising the amino acid sequence from the 1st amino acid Met to the 237th amino acid Ser of SEQ ID NO: 5, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added, and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 237th amino acid Ser of SEQ ID NO: 5;

- (4) a protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 7, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added, and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 7;
- (5) a protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 19, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added, and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 19;
- (6) a protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 21, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added, and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 583th amino acid Ser of SEQ ID NO: 21;
- (7) a protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 2, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 361st amino acid Ser of SEQ ID NO: 1;
- (8) a protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 4, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 3;
- (9) a protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 6, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 237th amino acid Ser of SEQ ID NO: 5;
- (10) a protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 8, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 7;
- (11) a protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 20, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 19;
- (12) a protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 22, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 21;
- (13) a fusion protein comprising the protein of any one of (1) to (12) and another peptide or polypeptide;
- (14) a DNA encoding the protein of any one of (1) to (13);
- (15) a vector comprising the DNA of (14);

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- (16) a transformant harboring the DNA of (14) in an expressible manner,
- (17) a method of producing the protein of any one of (1) to (13), comprising the step of culturing the transformant of (16);
- (18) a method of screening a compound that binds to the protein of any one of (1) to (13) comprising the steps of,
 - (a) contacting a test sample with the protein of any one of (1) to (13), and
 - (b) selecting a compound that comprises an activity to bind to the protein of any one of (1) to (13);
- (19) an antibody that specifically binds to the protein of any one of (1) to (12);
 - (20) a method of detecting or measuring the protein of any one of (1) to (13) comprising the steps of contacting a test sample presumed to contain said protein with the antibody of (19), and detecting or measuring the formation of the immune complex between the antibody and the protein; and
 - (21) a DNA specifically hybridizing to a DNA comprising the nucleotide sequence of any one of SEQ ID NOs: 2, 4, 6, 8, 20, and 22 to 27, and comprising at least 15 nucleotides.

[0020] The present invention relates to the novel hemopoietin receptor "NR8." 5'-RACE and 3'-RACE analyses, NR8 genome sequence analysis, and plaque screening analysis revealed the presence of NR8α, NR8β, and NR8γ. The structures of these NR8 genes are shown in Fig. 13. Among the NR8 genes, NR8β is an alternative splicing product lacking the 5th exon, and can encode two different proteins, a soluble protein in which the CDS ends with a stop codon on the 6th exon that results from a frame shift following direct coupling to the 4th exon, and a membrane-bound protein lacking the signal sequence starting from the ATG upon the 4th exon.

[0021] Since the soluble protein comprises the same sequence as NR8α up to the 4th exon, it may function as a

soluble receptor. On the other hand, NR8 γ encodes a protein containing a 177 amino acid insertion derived from the NR8 9^{th} intron close to the C terminus of the NR8 α as a result of selective splicing.

[0022] Both NR8 α and NR8 γ encode transmembrane-type hemopoietin receptors. Among the sequences conserved between other hemopoietin receptors that are thought to be involved in signal transduction, a motif resembling Box 1 exists in the intracellular domain of NR8 α and NR8 γ adjacent to the cell membrane. Though low in the degree of conservation, a sequence that is similar to Box 2 also exists, and therefore, NR8 is thought to be a type of receptor in which the signal is transduced by a homodimer.

[0023] The amino acid sequences of the NR8 proteins included in the proteins of the present invention are shown in SEQ ID No: 1 (NR8α), SEQ ID NO: 3 (soluble NR8β), SEQ ID NO: 5 (membrane-bound NR8β), and SEQ ID NO: 7 (NR8γ), and the nucleotide sequences of cDNA encoding these proteins are shown in SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, and SEQ ID NO: 8, respectively.

[0024] Northern blot analysis for the spleen, thymus, peripheral leucocytes, and lung showed two to three bands in the 5kb and 3 to 4kb regions. Similar sized bands were observed for cell lines HL60 and Raji also, but no expression was seen for other tumor cell lines (HeLa, SW480, A549, G361) and leukemia cell lines (K562, MOLT4).

[0025] The above results suggest that NR8 is specifically expressed on hemopoietic cell lines, especially on granulocytic lines, and B cell lines.

[0026] The above NR8 protein is expected to be applied in medicine. NR8 is expressed in fetal liver, spleen, thymus, and some leukemic cell lines, suggesting the possibility that it might be a receptor of an unknown hemopoietic factor. Therefore, NR8 protein would be a useful material for obtaining this unknown hemopoietic factor.

[0027] Furthermore, it is possible that NR8 is specifically expressed in limited cell populations within these hemopoietic tissues, and therefore, anti NR8 antibody may be useful as a means of separating these cell populations. Thus separated cell populations can be applied for cell transplant therapy. Anti NR8 antibody is also expected to be applied for the diagnosis and treatment of leukemic diseases represented by leukemia.

[0028] On the other hand, the soluble protein including the extracellular domain of NR8 protein, or NR8β, a splicing variant of NR8, may be applied as a decoy-type receptor that is an inhibitor of the NR8 ligand, and is anticipated to be applied in the treatment of diseases in which NR8 is involved, starting with leukemia.

[0029] The inventors also isolated mouse NR8 cDNA corresponding to the human-derived NR8 cDNA above-mentioned, by using the xenogeneic cross PCR cloning method. The amino acid sequences of the proteins named mouse NR8, which are included in the protein of the present invention are shown in SEQ ID NO: 19 (soluble mouse NR8β) and SEQ ID NO: 21 (mouse NR8γ), and the nucleotide sequences of the cDNA encoding these proteins are shown in SEQ ID NO: 20 and SEQ ID NO: 22, respectively.

[0030] As a result of structural analysis of the obtained mouse cDNA clones, alike human-derived NR8, the presence of mouse NR8γ encoding a transmembrane receptor protein comprising 538 amino acids and mouse NR8β encoding a secretory soluble receptor-like protein comprising 144 amino acids which were confirmed by the difference of transcripts derived the splice variant, was confirmed. When the amino acid sequences encoded by these receptor genes were compared between human and mouse, a high homology of 98.9% was observed for NR8γ, while a homology of 97.2% was seen for NR8β as well.

[0031] Northern blot analysis and RT-PCR analysis showed that although there were deviations in expression levels, mouse NR8 gene expression was seen in all organs analyzed, and seemed to be widely distributed compared to human NR8, for which a strong expression was seen only in immunocompetent and hemopoietic tissues. This also suggests the possibility that molecular functions of mouse NR8 may span a broad range of physiological regulatory mechanisms of the body.

[0032] The present invention also encompasses a protein that is functionally equivalent to the above-mentioned human or mouse NR8 protein. Herein "functionally equivalent" means having an equivalent biological activity to the above-mentioned NR8 proteins. Hemopoletic factor receptor protein activity can be given as an example of a biological activity. Such proteins can be obtained by the method of introducing a mutation to the amino acid sequence of a protein. For example, site-specific mutagenesis using a synthetic oligonucleotide primer, can be used to introduce a desired mutation into the amino acid sequence of a protein (Kramer, W. and Fritz, H.J., Methods in Enzymol., 1987, 154, 350-367). This could also be done by a PCR-mediated site-specific mutagenesis system (GIBCO-BRL). Using these methods, the amino acid sequence of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 19, or SEQ ID NO: 21 can be modified to obtain a protein functionally equivalent to the NR8 protein,in which one or more amino acids in the amino acid sequence of the protein have been deleted, added, and/or substituted by another amino acid without affecting the biological activity of the protein.

[0033] As a protein functionally equivalent to the NR8 protein of the invention, the following are given: one in which one or two or more, preferably, two to 30, more preferably, two to ten amino acids are deleted in any one of the amino acid sequences of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 19, or SEQ ID NO: 21; one in which one or two or more, preferably, two to 30, more preferably, two to ten amino acids have been added into any one of the amino acid sequences of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, or SEQ ID NO: 7; or one in which

one or two or more, preferably, two to 30, more preferably, two to ten amino acids have been substituted with other amino acids in any one of the amino acid sequences of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, or SEQ ID NO: 7.

[0034] It is already known that a protein comprising a modified amino acid sequence of a certain amino acid sequence in which one or more amino acid residues have been deleted, added, and/or substituted with another amino acid, still maintains its biological activity (Mark, D. F. et al., Proc. Natl. Acad. Sci. USA, 1984, 81, 5662-5666; Zoller, M. J. & Smith, M., Nucleic Acids Research, 1982, 10, 6487-6500; Wang, A. et al., Science, 224, 1431-1433; Dalbadie-McFarland, G. et al., Proc. Natl. Acad. Sci. USA, 1982, 79, 6409-6413).

[0035] For example, a fusion protein can be given as a protein in which one or more amino acid residues have been added to the NR8 protein of the present invention. A fusion protein is made by fusing the NR8 protein of the present invention with another peptide or protein and is encompassed in the present invention. A fusion protein can be prepared by ligating DNA encoding the NR8 protein of the present invention with DNA encoding another peptide or protein so as the frames match, introducing this into an expression vector, and expressing the fusion gene in a host. Methods commonly known can be used for preparing such a fusion gene. There is no restriction as to the other peptide or protein that is fused to the protein of this invention.

[0036] For example, FLAG (Hopp, T.P. et al., Biotechnology, 1988, 6, 1204-1210), 6x His constituting six histidine (His) residues, 10x His, Influenza agglutinin (HA), human c-myc fragment, VSV-GP fragment, p18HIV fragment, T7-tag, HSV-tag, E-tag, SV40T antigen fragment, lck tag, α-tubulin fragment, B-tag, Protein C fragment, and such well-known peptides can be used. Examples of proteins are, glutathione-S-transferase (GST), influenza agglutinin (HA), immunoglobulin constant region, β-galactosidase, maltose-binding protein (MBP), etc. Commercially available DNAs encoding these may also be used to prepare fusion proteins.

[0037] The protein of the invention can also be encoded by a DNA that hybridizes under stringent conditions to a DNA comprising any one of the nucleotide sequences of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 20, and SEQ ID NO: 22 to 27. Such a protein also includes a protein functionally equivalent to the above-mentioned NR8 protein. Stringent conditions can be suitably selected by one skilled in the art, and for example, low-stringent conditions can be given. Low-stringent conditions are, for example, 42°C, 2x SSC, and 0.1% SDS, and preferably, 50°C, 2x SSC, and 0.1% SDS. More preferable are highly stringent conditions, for example, 65°C, 2x SSC, and 0.1% SDS. Under these conditions, the higher the temperature is raised, the higher the homology of the obtained DNA will be.

[0038] The present invention also includes a protein that is functionally equivalent to the above NR8 protein, which has also a homology with a protein comprising any one of the amino acid sequences of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 19, or SEQ ID NO: 21. A protein having a homology means, a protein having at least 70%, preferably at least 80%, more preferably at least 90%, even more preferably, at least 95% homology to any one of the amino acid sequences of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, and SEQ ID NO: 7. The homology of a protein can be determined by the algorithm in "Wilbur, W.J. and Lipman, D.J. Proc. Natl. Acad. Sci. USA, 1983, 80, 726-730."

[0039] In the protein of the invention, the amino acid sequence, molecular weight, isoelectric point, the presence or absence of the sugar chain, and its form differ according to the producing cells, host, or purification method described below. However, as long as the obtained, protein comprises a hemopoietic factor receptor protein activity, it is included in the present invention.

[0040] For example, if the protein of the present invention is expressed in prokaryotic cells such as *E. coli*, a methionine residue is added at the N-terminus of the amino acid sequence of the expressed protein. If the protein of the present invention is expressed in eukaryotic cells such as mammalian cells, the N-terminal signal sequence is removed. The protein of the present invention includes these proteins.

[0041] For example, as a result of analyzing the protein of the invention based on the method in "Von Heijne, G., Nucleic Acids Research, 1986, 14, 4683-4690," it was presumed that the signal sequence is from the 1st Met to the 19th Gly in the amino acid sequence of SEQ ID NO: 1. Therefore, the present invention encompasses a protein comprising the sequence from the 20th Cys to 361st Ser in the amino acid sequence of SEQ ID NO: 1.

[0042] To produce the protein of the invention, the obtained DNA is incorporated into an expression vector in a manner that the DNA is expressible under the regulation of an expression regulatory region, for example, an enhancer or promoter. Next, host cells are transformed by this expression vector to express the protein.

[0043] Specifically, the protein can be produced as follows. When mammalian cells are used, DNA comprising a commonly used useful promoter/enhancer, DNA encoding the protein of the invention, and the poly A signal that is functionally bound to the 3' side downstream of the protein-encoding DNA, or a vector containing it, is constructed. For example, as the promoter/enhancer, human cytomegalovirus immediate early promoter/enhancer can be given.

[0044] Also, as other promoters/enhancers that can be used for protein expression, viral promoters/enhancers of retroviruses, polyomaviruses, adenoviruses, simian virus 40 (SV40), and such, and promoters/enhancers derived from mammalian cells, such as that of human elongation factor 1α (HEF1α) can be used.

[0045] For example, a protein can be easily expressed by following the method of Mulligan et al. (Nature, 1979, 277,

108) when using the SV40 promoter/enhancer, and the method of Mizushima et al. (Nucleic Acids Res., 1990, 18, 5322) when using the HEF1 α promoter/enhancer.

[0046] When using *E. coli*, well-used useful promotors, the signal sequence for polypeptide secretion, and genes to be expressed, may be functionally bound to express the desired gene. For example, lacZ promoter and araB promoter may be used as promotors. When using the lacZ promoter, the method of Ward et al. (Nature, 1098, 341, 544-546; FASEB J., 1992, 6, 2422-2427), and when using the araB promoter, the method of Better et al. (Science, 1988, 240, 1041-1043) may be followed.

[0047] When producing the protein into the periplasm of *E. coli*, the pelB (Lei, S. P. et al., J. Bacteriol., 1987, 169, 4379) signal sequence may be used as a protein secretion signal.

[0048] A replication origin derived from SV40, polyomavirus, adenovirus, bovine papilomavirus (BPV), and such may be used. To amplify gene copies in host cell lines, the expression vector may include an aminoglycoside transferase (APH) gene, thymidine kinase (TK) gene, E.coli xanthine guanine phosphoribosyl transferase (Ecogpt) gene, dihydrofolate reductase (dhfr) gene, and such as a selective marker.

[0049] The expression vector used to produce the protein of the invention may be any, as long as it's an expression vector that is suitably used for the present invention. Mammalian expression vectors, for example, pEF and pCDM8; insect-derived expression vectors, for example, pBacPAK8; plant-derived expression vectors, for example, pBacPAK8; plant-derived expression vectors, for example, pHSV, pMV, and pAdexLcw; retrovirus-derived expression vectors, for example, pNV11 and SP-Q01; Bacillus subtilisderived expression vectors, for example, pPL608 and pKTH50; E. coli-derived expression vectors, for example, pQE, pGEAPP, pGEMEAPP, and pMALp2 can be given as expression vectors of this invention.

[0050] Not only vectors that produce the protein of the invention *in vivo* and *in vitro*, but also those that are used for gene therapy of mammals, for example humans, are also included as vectors of the present invention.

[0051] When introducing the expression vector of the present invention constructed above into a host cell, well-known methods, for example the calcium phosphate method (Virology, 1973, 52, 456-467), electroporation (EMBO J., 1982, 1, 841-845), and such may be used.

[0052] In the present invention, an arbitrary production system may be used to produce the protein. *In vitro* and *in vivo* production systems are known as production systems for producing proteins. Production systems using eukaryotic cells and prokaryotic cells may be used as *in vitro* production systems.

[0053] When using eukaryotic cells, production systems using, for example, animal cells, plant cells, and fungal cells are known. As animal cells used, for example, mammalian cells such as CHO (J. Exp. Med., 1995, 108, 945), COS, myeloma, baby hamster kidney (BHK), HeLa, or Vero, amphibian cells such as *Xenopus* oocytes (Valle, et al., Nature, 1981, 291, 358-340), insect cells such as sf9, sf21, or Tn5, are known. As CHO cells, especially DHFR genedeficient CHO cell, dhfr-CHO (Proc. Natl. Acad. Sci. USA, 1980, 77, 4216-4220), and CHO K-1 (Proc. Natl. Acad. Sci. USA, 1968, 60, 1275) can be suitably used.

[0054] Nicotiana tabacum-derived cells are well known as plant cells, and these can be callus cultured. As fungal cells, yeasts such as the Saccharomyces genus, for example, Saccharomyces cerevisiae, filamentous bacteria such as the Aspergillus genus, for example, Aspergillus niger are known.

[0055] Bacterial cells may be used as prokaryotic production systems. As bacterial cells, *E. coli* and *Bacillus subtilis* are known.

[0056] Proteins can be obtained by transforming these cells with the objective DNA, and culturing the transformed cells in vitro according to well-known methods. For example, DMEM, MEM, RPMI1640, and IMDM can be used as culture media. At that instance, fetal calf serum (FCS) and such serum supplements may be added in the above media, or a serum-free culture medium may be used. The pH is preferably about 6 to 8. Culture is usually done at about 30°C to 40°C, for about 15 to 200 hr, and medium changes, aeration, and stirring are done as necessary.

[0057] On the other hand, production systems using animals and plants may be given as in vivo production systems. The objective gene is introduced into the plant or animal, and the protein is produced within the plant or animal, and recovered. "Host" as used in the present invention encompasses such animals and plants as well.

[0058] When using animals, mammalian and insect production systems can be used. As mammals, goats, pigs, sheep, mice, and cattle may be used (Vicki Glaser, SPECTRUM Biotechnology Applications, 1993). Transgenic animals may also be used when using mammals.

[0059] For example, the objective DNA is inserted within a gene encoding a protein produced intrinsically into milk, such as goat β casein, to prepare a fusion gene. The DNA fragment containing the fusion gene is injected into a goat's embryo, and this embryo is implanted in a female goat. The protein is collected from the milk of the transgenic goats produced from the goat that received the embryo, and descendents thereof. To increase the amount of protein-containing milk produced from the transgenic goat, a suitable hormone/hormones may be given to the transgenic goats (Ebert, K.M. et al., Bio/Technology, 1994, 12, 699-702).

[0060] Silk worms may be used as insects. When using the silk worm, it is infected with a baculovirus to which the objective DNA has been inserted, and the desired protein is obtained from the body fluids of the silk worm (Susumu, M.

et al., Nature, 1985, 315, 592-594).

[0061] When using plants, for example, tobacco can be used. In the case of tobacco, the objective DNA is inserted into a plant expression vector, for example pMON 530, and this vector is introduced into a bacterium such as *Agrobacterium tumefaciens*. This bacterium is infected to tobacco, for example *Nicotiana tabacum*, to obtain the desired polypeptide from tobacco leaves (Julian, K.-C. Ma et al., Eur. J. Immunol., 1994, 24, 131-138).

[0062] The thus-obtained protein of the invention is isolated from within and without cells, or from hosts, and can be purified as a substantially pure homogenous protein. The separation and purification of the protein is not limited to any specific method and can be done using ordinary separation and purification methods used to purify proteins. For example, chromatography, filtration, ultrafiltration, salting out, solvent precipitation, solvent extraction, distillation, immunoprecipitation, SDS-polyacrylamide gel electrophoresis, isoelectric focusing, dialysis, recrystalization, and such may be suitably selected, or combined to separate/purify the protein.

[0063] As chromatographies, for example, affinity chromatography, ion exchange chromatography, hydrophobic chromatography, gel filtration, reversed-phase chromatography, adsorption chromatography, and such can be exemplified (Strategies for Protein Purification and Characterization: A Laboratory Course Manual. Ed Daniel R. Marshak et al., Cold Spring Harbor Laboratory Press, 1996). These chromatographies can be done by liquid chromatography such as HPLC, FPLC, and the like. The present invention encompasses proteins highly purified by using such purification methods.

[0064] Proteins can be arbitrarily modified, or peptides may be partially excised by treating the proteins with appropriate modification enzymes prior to or after the purification. Trypsin, chymotrypsin, lysyl endopeptidase, protein kinase, glucosidase, and such are used as protein modification enzymes.

The present invention includes a partial peptide comprising the active center of a protein comprising any one of the amino acid sequences of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 19, and SEQ ID NO: 21. A partial peptide of the protein of the present invention is, for example, a partial peptide of the molecules of the protein, which contains one or more regions of the hydrophilic region and hydrophobic region presumed by hydrophobicity plot analysis. These partial peptides may contain the whole hydrophobic region or a part of it. For example, soluble proteins and proteins comprising extracellular regions of the protein of the invention, are also encompassed in the invention.

[0066] The partial peptides of the protein of the invention may be produced by genetic engineering techniques, well-known peptide synthesizing methods, or by excising the protein of the invention by a suitable peptidase. As peptide synthesizing methods, the solid-phase synthesizing method, and the liquid-phase synthesizing method may be used.

[0067] The present invention also relates to a DNA encoding the protein of the invention. A cDNA encoding the protein of the invention may be obtained by, for example, screening a human cDNA library using the probe described herein.

[0068] Using the obtained cDNA or cDNA fragment as a probe, cDNA can also be obtained from other cells, tissues, organs, or species by further screening cDNA libraries. cDNA libraries may be prepared by, for example, the method of Sambrook, J. et al., Molecular Cloning, Cold Spring Harbor Laboratory Press (1989), or commercially available cDNA libraries may be used.

[0069] By determining the nucleotide sequence of the obtained cDNA, the translation region encoded by it can be determined, and the amino acid sequence of the protein of the present invention can be obtained. Furthermore, genomic DNA can be isolated by screening the genomic DNA library using the obtained cDNA as a probe.

[0070] Specifically, this can be done as follows. First, mRNA is isolated from cells, tissues, and organs expressing the protein of the invention. For this mRNA isolation, whole RNA is prepared using well-known methods, for example, guanidine ultracentrifugation method (Chirgwin, J.M. et al., Biochemistry, 1979, 18, 5294-5299), the AGPC method (Chomczynski, P. and Sacchi, N., Anal. Biochem., 1987, 162, 156-159), and such, and purified using the mRNA Purification Kit (Pharmacia), etc. mRNA may be directly prepared using the QuickPrep mRNA Purification Kit (Pharmacia).

[0071] cDNA is synthesized using reverse transcriptase from the obtained mRNA. cDNA can be synthesized using the AMV Reverse Transcriptase First-strand cDNA Synthesis Kit (SEIKAGAKU CORPORATION), etc. Also, cDNA synthesis and amplification may also be done using the probe described herein by following the 5'-RACE method (Frohman, M.A. et al., Proc. Natl. Acad. Sci. U.S.A., 1988, 85, 8998-9002; Belyavsky, A. et al., Nucleic Acids Res., 1989, 17, 2919-2932) using the polymerase chain reaction (PCR) and the 5'-Ampli FINDER RACE KIT (Clontech).

[0072] The objective DNA fragment is prepared from the obtained PCR product and ligated with vector DNA. Thus, a recombination vector is created, introduced into *E.coli*, etc. and colonies are selected to prepare the desired recombination vector. The nucleotide sequence of the objective DNA may be verified by known methods, for example, the dideoxy nucleotide chain termination method.

[0073] In the DNA of the invention, a sequence with a higher expression efficiency can be designed by considering the codon usage frequency of hosts used for the expression (Grantham, R. et al., Nucleic Acids Research, 1981, 9, p43-p74). The DNA of the invention may also be modified using commercially available kits and known methods. For example, digestion by restriction enzymes, insertion of synthetic oligonucleotides and suitable DNA fragments, addition

of linkers, insertion of a start codon (ATG) and/or stop codon (ATT, TGA, or TAG), and such can be given.

[0074] The DNA of the present invention encompasses DNA comprising the nucleotide sequence from the 441st nucleotide A to the 1523rd nucleotide C in the nucleotide sequence of SEQ ID NO: 2, DNA comprising the nucleotide sequence from the 441st nucleotide A to the 872nd nucleotide A in the nucleotide sequence of SEQ ID NO: 4, DNA comprising the nucleotide sequence from the 659th nucleotide A to the 1368th nucleotide C in the nucleotide sequence of SEQ ID NO: 6, DNA comprising the nucleotide sequence from the 441st nucleotide A to the 2054th nucleotide C in the nucleotide sequence of SEQ ID NO: 8, DNA comprising the nucleotide sequence from the 439th nucleotide A to the 870th nucleotide A in the nucleotide sequence of SEQ ID NO: 20, and DNA comprising the nucleotide sequence from the 439th nucleotide A to the 2052nd nucleotide C in the nucleotide sequence of SEQ ID NO: 22.

[0075] The DNA of the present invention encompasses DNA that hybridizes under stringent conditions to the DNA comprising any one of the nucleotide sequences of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 20, and SEQ ID NO: 22 to 27, which also includes a DNA encoding a protein functionally equivalent to the NR8 protein.

[0076] Stringent conditions can be suitably selected by one skilled in the art, and for example, low-stringent conditions can be given. Low-stringent conditions are, for example, 42°C, 2x SSC, and 0.1% SDS, and preferably 50°C, 2x SSC, and 0.1% SDS. More preferable are highly stringent conditions, for example, 65°C, 2x SSC, and 0.1% SDS. Under these conditions, the higher the temperature is raised, the higher the homology of the obtained DNA will be. The above DNA is preferably natural DNA such as cDNA and chromosomal DNA.

[0077] As shown in Examples, the mRNA of the gene hybridizing to cDNA encoding the protein of the invention was distributed in various human tissues. Therefore, the above-mentioned natural DNA may be, for example, genomic DNA and cDNA derived from tissues in which the mRNA that hybridizes to the cDNA encoding the protein of the invention is detected in Examples. The DNA encoding the protein of the invention may be cDNA, genomic DNA, or synthetic DNA. [0078] The protein of the invention is useful in screening a compound that binds to it. Namely, the protein of the invention is used in the screening method that comprises the steps of contacting a test sample expected to contain a compound that binds to the protein of the invention with the protein of the invention, and selecting the compound that comprises an activity to bind to the protein of the invention.

[0079] As methods for screening a compound that comprises an activity to bind to the protein of the invention, numerous methods usually used by those skilled in the art can be employed. The protein of the invention that is used for the screening of the invention may be a recombinant, natural, or partial peptide. A compound comprising an activity to bind to the protein of the invention may be a protein comprising a binding activity, or it may be a chemically synthesized compound having a binding activity.

[0080] As a test sample that is used in the screening method of the present invention, for example, peptides, purified or crudely purified proteins, non-peptide compounds, synthetic compounds, microbial fermentation products, extracts of marine organisms, plant extracts, cell extracts, animal tissue extracts, and such can be given. These test samples may be novel compounds, or well-known compounds.

[0081] A protein that binds to the protein of the invention can be screened by, for example, using the West-western blotting method (Skolnik, E.Y. et al., Cell, 1991, 65, 83-90). cDNA is isolated from cells, tissues, and organs presumed to express the protein binding to the protein of the invention, this is inserted into phage vectors, for example, λgt11, ZAPII, and such, to make a cDNA library, expressed on a plate containing a culture medium, the proteins expressed are fixed on a filter, this filter is reacted with the labeled, purified protein of the invention, and plaques expressing the protein bound to the protein of the invention are detected by the labels. As methods to label the protein of the invention, the method that uses the binding ability of avidin and biotin, the method of using an antibody that specifically binds to the protein of the invention or the peptide or polypeptide fused to the protein of the invention, the method of using radioisotopes, or fluorescence, and such can be given.

[0082] A ligand that binds specifically to the protein of the invention can be screened by, preparing a chimeric receptor by ligating the extracellular domain of the protein of the invention with the intracellular domain containing the transmembrane domain of a hemopoletin receptor protein comprising a known signal transduction ability, expressing this chimeric receptor on the cell surface of a suitable cell line, preferably, a cell line that can survive and proliferate under the presence of a suitable growth factor (a growth factor-dependent cell line), and culturing the cell line by adding a material that is expected to contain various growth factors, cytokines, or hemopoletic factors. This method uses the fact that the above-mentioned growth factor-dependent cell line survives and proliferates only when a ligand that specifically binds to the extracellular domain of the protein of the invention exists within the test material. Known hemopoletic receptors are, for example, the thrombopoletin receptor, erythropoletin receptor, G-CSF receptor, gp130, etc. However, the partner of the chimeric receptor used in the screening of the invention is not limited to these known hemopoletic receptors, and any may be used as long as a structure needed for the signal transduction activity is contained in the cytoplasmic domain. Growth factor-dependent cell lines are for example, IL-3-dependent cell lines starting with BaF3 and FDC-P1.

[0083] As a ligand that specifically binds to the protein of the invention, the possibility of not only soluble proteins,

but also cell membrane-binding proteins can be envisaged, though rare. In such cases, screening can be done by labeling the protein containing only the extracellular domain of the protein of the invention, or a fusion protein in which the partial sequence of another soluble protein has been added to this extracellular domain, and measuring the binding with cells expected to express the ligand. As examples of proteins containing only the extracellular domain of the protein of the invention, for example, a soluble receptor protein artificially made by inserting a stop codon to the N terminal side of the transmembrane domain, or NR8 β soluble protein may be used. On the other hand, as a fusion protein in which the partial sequence of another soluble protein has been added to the extracellular domain of the protein of the invention, for example, proteins prepared by adding immunoglobulin Fc site, FLAG peptide, etc. to the C terminus of the extracellular domain can be used. These soluble labeled proteins can be used in the detection in the above-described West-western blotting method.

[0084] A protein that binds to the protein of the invention can be screened by using the two-hybrid system (Fields, S. and Sternglanz, R., Trends. Genet., 1994, 10, 286-292).

[0085] In the two-hybrid system, an expression vector containing DNA encoding the fusion protein between the protein of the invention and one subunit of a heterodimeric transcriptional regulatory factor, and an expression vector containing DNA made by ligating DNA encoding the other subunit of the heterodimeric transcriptional regulatory factor and a desired cDNA used as a test sample are introduced into cells and expressed. If the protein encoded by the cDNA binds with the protein of the invention and the transcriptional regulatory factor forms a heterodimer, a reporter gene constructed in the cell beforehand will be expressed. Therefore, a protein binding to the protein of the invention can be selected by detecting or measuring the expression level of the reporter gene.

[0086] Specifically, the DNA encoding the protein of the invention and the gene encoding the DNA binding domain of LexA are ligated so as the frames match to prepare an expression vector. Next, the desired cDNA and the gene encoding GAL4 transcription activation domain are ligated to prepare an expression vector.

[0087] Cells into which the HIS3 gene has been incorporated (the transcription of HIS3 gene is regulated by the promoter having a LexA binding motif) are transformed by the above two-hybrid system expression plasmids, and then incubated on a histidine-free synthetic culture medium. Herein, cells only grow when a protein interaction is present. Thus, the increase in reporter gene expression can be examined by the growth rate of the transformant.

[0088] Other than the HIS3 gene, for example, the luciferase gene, plasminogen activator inhibitor type1 (PAI-1) gene, ADE2 gene, LacZ gene, CDC25H gene, and such can be used as reporter genes.

[0089] The two-hybrid system may be constructed according to the usual methods, or a commercially available kit may be used. As commercially available two-hybrid system kits, the MATCHMARKER Two-Hybrid System, Mammalian MATCHMARKER Two-Hybrid Assay Kit (both by CLONTEC), HybriZAP Two-Hybrid Vector System (Stratagene), and CytoTrap two-hybrid system (Stratagene) can be given.

[0090] A protein binding to the protein of the invention can be screened by affinity chromatography. Namely, the protein of the invention is immobilized onto a carrier of an affinity column, and a test sample presumed to express a protein binding to the protein of the invention is applied to the column. As this test sample, a cell culture supernatant, cell extract, cell lysate, and such may be used. After applying the test sample, the column is washed to obtain the protein binding to the protein of the invention.

[0091] The compound isolated by the screening method of the invention is a candidate drug for promoting or inhibiting the activity of the protein of the invention. The compound obtained by using the screening method of the invention encompasses a compound resulting from modifying the compound having an activity to bind to the protein of the invention by adding, deleting, and/or replacing a part of the structure.

[0092] When using the compound obtained by the screening method of the invention as drugs for humans and other mammals such as, mice, rats, guinea pigs, rabbits, chicken, cats, dogs, sheep, pigs, cattle, monkeys, sacred baboons, and chimpanzees, the drug may be administered using ordinary means.

[0093] For example, according to the need, the drugs can be taken orally as sugar-coated tablets, capsules, elixirs, and microcapsules, or parenterally in the form of injections of sterile solutions or suspensions with water or any other pharmaceutically acceptable liquid. For example, the compounds comprising the activity to bind to the protein of the invention can be mixed with physiologically acceptable carriers, flavoring agents, excipients, vehicles, preservatives, stabilizers, and binders, in a unit dose form required for generally accepted drug implementation. The amount of active ingredients in these preparations makes a suitable dosage within the indicated range acquirable.

[0094] Examples of additives that can be mixed to tablets and capsules are, binders such as gelatin, corn starch, tragacanth gum, and arabic gum; excipients such as crystalline cellulose; swelling agents such as cornstarch, gelatin, and alginic acid; lubricants such as magnesium stearate; sweeteners such as sucrose, lactose, or saccharin; and flavoring agents such as peppermint, Gaultheria adenothrix oil, and cherry. When the unit dosage form is a capsule, a liquid carrier, such as oil, can also be included in the above additives. Sterile compositions for injections can be formulated following usual drug implementations using vehicles such as distilled water used for injections.

[0095] For example, physiological saline and isotonic liquids including glucose or other adjuvants, such as D-sorbitol, D-mannose, D-mannitol, and sodium chloride, can be used as aqueous solutions for injections. These can be used

in conjunction with suitable solubilizers, such as alcohol, specifically ethanol, polyalcohols such as propylene glycol and polyethylene glycol, non-ionic surfactants, such as Polysorbate 80 (TM) and HCO-50.

[0096] Sesame oil or soy-bean oil can be used as a oleaginous liquid and may be used in conjunction with benzyl benzoate or benzyl alcohol as a solubilizer; may be formulated with a buffer such as phosphate buffer and sodium acetate buffer; a pain-killer such as procaine hydrochloride; a stabilizer such as benzyl alcohol and phenol; and an anti-oxidant. The prepared injection is usually filled into a suitable ampule.

[0097] Although the dosage of the compound that has the activity to bind to the protein of the invention varies according to symptoms, the daily dose is generally about 0.1 to about 100 mg, preferably about 1.0 to about 50 mg, and more preferably about 1.0 to about 20 mg, when administered orally to an adult (body weight 60 kg).

[0098] When given parenterally, the dose differs according to the patient, target organ, symptoms, and method of administration, but the daily dose is usually about 0.01 to about 30 mg, preferably about 0.1 to about 20 mg and more preferably about 0.1 to about 10 mg for an adult (body weight 60 kg) when given as an intravenous injection. Also, in the case of other animals too, it is possible to administer an amount converted to 60 kg of body-weight.

[0099] The antibody of the present invention can be obtained as a monoclonal antibody or a polyclonal antibody using well-known methods.

[0100] The antibody that specifically binds to the protein of the invention can be prepared by using the protein of the invention as a sensitizing antigen for immunization according to usual immunizing methods, fusing the obtained immunized cells with known parent cells by ordinary cell fusion methods, and screening for antibody producing cells using the usual screening techniques.

[0101] Specifically, a monoclonal or polyclonal antibody that binds to the proteins of the invention may be prepared as follows.

[0102] For example, the protein of the invention that is used as a sensitizing antigen for obtaining the antibody is not restricted by the animal species from which it is derived, but is preferably a protein derived from mammals, for example, humans, mice, or rats, especially from humans. Proteins of human origin can be obtained by using the nucleotide sequence or amino acid sequence disclosed herein.

[0103] The protein that is used as a sensitizing antigen in the present invention can be a protein that comprises the biological activity of all the proteins described herein. Partial peptides of the proteins may also be used. As partial peptides of the proteins, for example, the amino (N) terminal fragment of the protein, and the carboxy (C) terminal fragment can be given. "Antibody" as used herein means an antibody that specifically reacts with the full-length or fragment of the protein.

[0104] A gene encoding the protein of the invention or a fragment thereof is inserted into a well-known expression vector, and after transforming the host cells described herein, the objective protein or a fragment thereof is obtained from within and without the host cell, or from the host using well-known methods, and this protein can be used as a sensitizing antigen. Also, cells expressing the protein, cell lysates, or chemically synthesized protein of the invention may be used as a sensitizing antigen.

[0105] The mammals that are immunized by the sensitizing antigen are not restricted, but it is preferable to select the animal by considering the adaptability with the parent cells used in cell fusion. Generally, an animal belonging to Rodentia, Lagomorpha, or Primates is used.

[0106] As animals belonging to Rodentia, for example, mice, rats, hamsters, and such are used. As animals belonging to Lagomorpha, for example rabbits, as Primates, for example monkeys, are used. As monkeys, monkeys of the infraorder Catarrhini (Old World Monkeys), for example, cynomolgus monkeys, rhesus monkeys, sacred baboons, chimpanzees, etc., are used.

[0107] To immunize animals with the sensitizing antigen, well-known methods may be used. For example, the sensitizing antigen is generally injected into mammals intraperitoneally or subcutaneously. Specifically, the sensitizing antigen is suitably diluted, suspended in physiological saline or phosphate-buffered saline (PBS), mixed with a suitable amount of a general adjuvant if desired, for example, with Freund's complete adjuvant, emulsified and injected into the mammal. Thereafter, the sensitizing antigen suitably mixed with Freund's incomplete adjuvant is preferably given several times every four to 21 days. A suitable carrier can also be used when immunizing an animal with the sensitizing antigen. After the immunization, the elevation in the serum antibody level is detected by usual methods.

[0108] Polyclonal antibodies against the protein of the invention can be obtained as follows. After verifying that the desired serum antibody level has been reached, blood is withdrawn from the mammal sensitized with the antigen. Serum is isolated from this blood using well-known methods. The serum containing the polyclonal antibody may be used as the polyclonal antibody, or according to needs, the polyclonal antibody-containing fraction may be further isolated from the serum.

[0109] To obtain monoclonal antibodies, after verifying that the desired serum antibody level has been reached in the mammal sensitized with the above-described antigen, immunocytes are taken from the mammal and used for cell fusion. At this instance, immunocytes that are preferably used for cell fusion are splenocytes. As parent cells fused with the above immunocytes, preferable are mammalian myeloma cells, more preferable are, myeloma cells that have

attained the feature of distinguishing fusion cells by agents.

[0110] For the cell fusion between the above immunocytes and myeloma cells, for example, the method of Milstein et al. (Galfre, G. and Milstein, C., Methods Enzymol., 1981, 73, 3-46) is basically well known.

[0111] The hybridoma obtained from cell fusion is selected by culturing in a usual selective culture medium, for example, HAT culture medium (hypoxanthine, aminopterin, thymidine-containing culture medium). The culture in this HAT medium is continued for a period sufficient enough for cells (non-fusion cells)other than the objective hybridoma to perish, usually from a few days to a few weeks. Next, the usual limiting dilution method is carried out, and the hybridoma producing the objective antibody is screened and cloned.

[0112] Other than the above method of obtaining a hybridoma by immunizing an animal other than humans with the antigen, a hybridoma producing the objective human antibodies comprising the activity to bind to proteins can be obtained by the method of sensitizing human lymphocytes, for example, human lymphocytes infected with the EB virus, with proteins, protein-expressing cells, or lysates thereof *in vitro*, fusing the sensitized lymphocytes with myeloma cells derived from human, for example U266, having the capacity of permanent cell division (Unexamined Published Japanese Patent Application (JP-A) No. Sho 63-17688).

[0113] Moreover, human antibody against the protein can be obtained using a hybridoma made by fusing myeloma cells with antibody-producing cells obtained by immunizing a transgenic animal comprising a repertoire of human antibody genes with an antigen such as a protein, protein-expressing cells, or a cell lysate thereof WO92/03918, WO93/2227, WO94/02602, WO94/25585, WO96/33735, and WO96/34096).

[0114] Other than producing antibodies by using hybridoma, antibody-producing immunocytes such as sensitized lymphocytes that are immortalized by oncogenes may also be used.

[0115] Such monoclonal antibodies can also be obtained as recombinant antibodies produced by using the gene engineering technique (for example, Borrebaeck, C.A.K. and Larrick, J.W., THERAPEUTIC MONOCLONAL ANTIBODIES, Published in the United Kingdom by MACMILLAN PUBLISHERS LTD, 1990). Recombinant antibodies are produced by cloning the encoding DNA from immunocytes such as hybridoma or antibody-producing sensitized lymphocytes, incorporating this into a suitable vector, and introducing this vector into a host to produce the antibody. The present invention encompasses such recombinant antibodies as well.

[0116] The antibody of the present invention may be an antibody fragment or a modified-antibody as long as it binds to the protein of the invention. For example, Fab, F(ab')₂, Fv, or single chain Fv in which the H chain Fv and the L chain Fv are suitably linked by a linker (scFv, Huston, J.S. et al., Proc. Natl. Acad. Sci. U.S.A., 1988, 85, 5879-5883) can be given as antibody fragments. Specifically, antibody fragments are produced by treating an antibody with an enzyme, for example, papain, pepsin, etc. or by constructing a gene encoding an antibody fragment, introducing this into an expression vector, and expressing this vector on suitable host cells (for example, Co, M.S. et al., J. Immunol., 1994, 152, 2968-2976; Better, M. and Horwitz, A.H., Methods Enzymol., 1989, 178, 476-496; Pluckthun, A. and Skerra, A., Methods Enzymol., 1989, 178, 497-515; Lamoyi, E., Methods Enzymol., 1986, 121, 662-663; Rousseaux, J. et al., Methods Enzymol., 1986, 121, 663-669; Bird, R.E. and Walker, B.W., Trends Biotechnol., 1991, 9, 132-137).

[0117] As a modified antibody, an antibody bound to various molecules such as polyethylene glycol (PEG) can be used. The present antibody encompasses such modified antibodies as well. To obtain such a modified antibody, chemical modifications are done to the obtained antibody. These methods are already established in the field.

[0118] The antibody of the invention may be obtained as a chimeric antibody comprising non-human antibody-derived variable region and a human antibody-derived constant region, or as a humanized antibody comprising non-human antibody-derived complementarity determining region (CDR), and human antibody-derived framework region (FR) and a constant region.

[0119] Antibodies thus obtained can be purified till uniform. The separation and purification methods for separating and purifying the antibody used in the present invention may be any method usually used for proteins, and is not in the least limited. Antibody concentration of the above mentioned antibody can be assayed by measuring the absorbance, or by the enzyme-linked immunosorbent assay (ELISA), etc.

[0120] Also, as methods that assay the antigen-binding activity of the antibody of the invention, ELISA, enzyme immunoassay (EIA), radio immunoassay (RIA), or fluorescent antibody method can be given. For example, when using ELISA, the protein of the invention is added to a plate coated with the antibody of the invention, and next, the objective antibody sample, for example, culture supernatants of antibody-producing cells, or purified antibodies are added. Then, secondary antibody recognizing the antibody, which is labeled by alkaline phosphatase and such enzymes, is added, the plate is incubated and washed, and absorbance is measured to evaluate the antigen-binding activity after adding an enzyme substrate such as p-nitrophenyl phosphate. As the protein, a protein fragment, for example, a fragment comprising a C terminus, or a fragment comprising an N terminus may be used. To evaluate the activity of the antibody of the invention, BlAcore (Pharmacia) may be used.

[0121] By using these methods, the antibody of the invention and a sample presumed to contain the protein of the invention are contacted, and the protein of the invention is detected or assayed by detecting or assaying the immune complex of the above-mentioned antibody and protein.

[0122] A method of detecting or assaying the protein of the invention is useful in various experiments using proteins as it can specifically detect or assay the proteins.

[0123] The present invention also encompasses a DNA specifically hybridizing to a DNA comprising a nucleotide sequence of any one of SEQ ID NOs: 2, 4, 6, 8, 20, and 22 to 27 or its complementary DNA, and comprising at least 15 nucleotides. Namely, a probe that can selectively hybridize to the DNA encoding the protein of the invention, or a DNA complementary to the above DNA, a nucleotide or nucleotide derivative, for example, antisense oligonucleotide, ribozyme, and such are included.

[0124] The present invention also encompasses an antisense oligonucleotide that hybridizes to any portion of any one of the nucleotide sequences shown in, for example, SEQ ID NOs: 2, 4, 6, 8, 20, and 22 to 27. This antisense oligonucleotide is preferably one against at least 15 continuous nucleotides in any one of the nucleotide sequences of SEQ ID NOs: 2, 4, 6, 8, 20, and 22 to 27. More preferable is the above-mentioned antisense oligonucleotide against the above-mentioned at least 15 continuous nucleotides containing a translation start codon.

[0125] Derivatives or modified products of antisense oligonucleotides can be used as antisense oligonucleotides. As such modified products, for example, lower alkyl phosphonate modifications such as methyl-phosphonate-type or ethyl-phosphonate-type, phosphorothioate or phosphoroamidate-modified products, etc. may be used.

[0126] The term "antisense oligonucleotide(s)" as used herein means, not only those in which the nucleotides corresponding to those constituting a specified region of a DNA or mRNA are entirely complementary, but also those having a mismatch of one or more nucleotides, as long as the DNA or mRNA and the oligonucleotide can selectively and stably hybridize with the nucleotide sequence of SEQ ID NO: 1.

[0127] "Selectively and stably hybridize" means that significant cross hybridization with DNA encoding other proteins does not occur under usual hybridization conditions, preferably under stringent hybridization conditions. Such DNAs are indicated as those having, in the "at least 15 continuous nucleotide" sequence region, a hornology of at least 70% or higher, preferably 80% or higher, more preferably 90% or higher, even more preferably 95% or higher nucleotide sequence homology. The algorithm stated herein can be used to determine homology. Such DNA is useful as a probe for detecting or isolating DNA encoding the protein of the invention, or as a primer for amplification as described in Examples below.

[0128] The antisense oligonucleotide derivative of the present invention acts upon cells producing the protein of the invention by binding to the DNA or mRNA encoding the protein to inhibit its transcription or translation, and to promote the degradation of mRNA, and has an effect of suppressing the function of the protein of the invention by suppressing the expression of the protein.

[0129] The antisense oligonucleotide derivative of the present invention can be made into an external preparation such as a liniment and a poultice by mixing with a suitable base material, which is inactive against the derivatives.

[0130] Also, as needed, the derivatives can be formulated into tablets, powders, granules, capsules, liposome capsules, injections, solutions, nose-drops, and freeze-dried agents by adding excipients, isotonic agents, solubilizers, stabilizers, preservatives, pain-killers, etc. These can be prepared using the usual methods.

[0131] The antisense oligonucleotide derivative is given to the patient by directly applying onto the ailing site, by injecting into a blood vessel, etc. so that it will reach the ailing site. An antisense-mounting material can also be used to increase durability and membrane-permeability. Examples are, liposome, poly-L lysine, lipid, cholesterol, lipofectin, or derivatives of these.

[0132] The dosage of the antisense oligonucleotide derivative of the present invention can be adjusted suitably according to the patient's condition and used in desired amounts. For example, a dose range of 0.1 to 100 mg/kg, preferably 0.1 to 50 mg/kg can be administered.

[0133] The antisense oligonucleotide derivative of the present invention is useful in inhibiting the expression of the protein of the invention, and therefore is useful in suppressing the biological activity of the protein of the invention. Also, expression-inhibitors comprising the antisense oligonucleotide derivative of the present invention are useful because of their capability to suppress the biological activity of the protein of the invention.

Brief Description of the Drawings

∞ [0134]

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Figure 1 is a schematic diagram showing the results of BlastX search where the query was 180 nucleotides of 40861-41040 including 40952-40966, the only probe sequence within the AC002303. *#*: For only NR8 the number was indicated by the nucleotide number. The underline of the NR8 sequence shows the portion corresponding to the exon. Other underlined sequences show identical amino acids.

· 1. 1. 李维 (12) 4.

Figure 2 is a schematic diagram showing the results of BlastX scanning of 180 nucleotides in both the 5' and 3' directions, where the search centered on the 180 nucleotides of 40861-41040 containing 40952-40966, the only probe sequence within the AC002303.

Figure 3 shows the electrophoresis results of the amplification done by the RT-PCR method for the combinations of SN1/AS1, SN1/AS2, SN2/AS1, and SN2/AS2 primers using human fetal liver and skeletal muscle cDNA as templates.

Figure 4 shows the electrophoretic results of the 5'-RACE method and 3'-RACE method using human fetal liver cDNA as the template.

Figure 5 shows the nucleotide sequence and the amino acid sequence of NR8 α cDNA. The arrows show the positions of primers used for RT-PCR. They are, SN1 (798-827), SN2 (894-923), AS2 (1055-1026), and AS1 (1127-1098) from the 5' side, in their order. For two bases at the 5' end of AS1, AC, which is derived from the genomic sequence, was used in place of CT.

Figure 6 is the continuation of Fig. 5 showing the nucleotide sequence and the amino acid sequence of NR8α cDNA.

Figure 7 shows the nucleotide sequence and the amino acid sequence of NR8β cDNA. Two possible open reading frames (ORF) are shown.

Figure 8 is the continuation of Fig. 7 showing the nucleotide sequence and the amino acid sequence of NR8ß cDNA.

Figure 9 shows the nucleotide sequence and the amino acid sequence of NR8γ cDNA. The 177 amino acids inserted by selective splicing are underlined.

Figure 10 is the continuation of Fig. 9 showing the nucleotide sequence and the amino acid sequence of NR8γ cDNA. The 177 amino acids inserted by selective splicing are underlined.

20 Figure 11 is the continuation of Fig. 10 showing the nucleotide sequence and the amino acid sequence of NR8y cDNA.

Figure 12 shows the results of Northern blot analysis of NR8 expression in each organ.

Figure 13 is a schematic diagram showing the structure of the NR8 gene. Other repetitives include, (CA)n, (CAGA)n, (TGGA)n, (TA)n, (TA)n, (GA)n, (GGAA)n, (CATG)n, (GAAA)n, MSTA, AT-rich, MLT1A1, LINE2, FLAM_C, MER63A, and MSTB.

Figure 14 is a schematic diagram showing the structure of expressible proteins constructed in the expression vector.

Figure 15 shows the results of cross PCR, in which the human NR8 primer set was used against a mouse cDNA library. As the size marker, 100 bp DNA Ladder (NEB#323-1L) was used.

Figure 16 shows a comparison between amino acid sequences of human and mouse NR8β. The amino acid sequences where the two coincide are shadowed. Also, cysteine residues conserved in other hemopoietin receptors are displayed in boldface type within the sequence.

Figure 17 shows a comparison between amino acid sequences of human and mouse NR8γ. The amino acid sequences where the two coincide are shadowed. Also, cysteine residues conserved in other hemopoietin receptors and the WSXWS-Box are displayed in boldface type within the sequence.

Figure 18 shows the results of NR8 gene expression analysis in each mouse organ using the RT-PCR method. The size marker, 100 bp DNA Ladder (NEB#323-1L), is shown on the either sides of the lane. A 320 bp target gene has been detected in all organs.

Figure 19 shows the results of NR8 gene expression analysis in each mouse organ using the Northern blotting method (left). An approximately 4.2 kb transcript was intensely detected in the testis only. Mouse β -actin was detected in the same blot as a positive control (right).

Best Mode for Carrying Out the Invention

45 [0135] The present invention shall be described in detail below with reference to examples, but is not be construed as being limited thereto.

Example 1: Two step Blast Search

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[0136] Probe sequences (256 types) comprising the tggag(t/c)nnntggag(t/c) (where n is an arbitrary nucleotide) as the oligonucleotide encoding the Trp-Ser-Xaa-Trp-Ser motif were designed. These sequences enable the detection of almost all known hemopoietin receptors, except for the EPO receptor, TPO receptor, and the mouse IL6 receptor. Using each sequence as the query, the GenBank nr database was searched using the BlastN (Advanced BlastN 2.0.4) program. Default values (Descriptions=100, Alignments=100) were used as parameters for the search, except for making the expectation value 100.

[0137] Since approximately 500 clones that completely matched the probe sequences were obtained as a result of the primary search, among these, a 180-residue nucleotide sequence of human genome-derived clones (cosmid, BAC, and PAC) containing the probe sequence in approximately the center was excised. Next, using this 180-residue nucle-

otide sequence as the query, the nr database was searched again using the BlastX (Advanced BlastX 2.0.4) program to search the homology of the amino acid sequence around the probe sequence with known hemopoietin receptors.

[0138] Default values were used as parameters for the search, except for making the expectation value 100. However, when extremely large number of hits were obtained (caused by the Alu sub family that is a high repetitive sequence), it was often difficult to observe hits for known hemopoietic receptors. Therefore, to maximize the sensitivity in such cases, a value of "Expect=1000, Descriptions=500, Alignments=500" was used.

[0139] As a result of the secondary search by BlastX, 28 clones hit one or more known hemopoietin receptors (Table 1 to Table 8).

Table 1

Probe "	Xaa	Accession No.	Hit site	Locas	Mark (contraction)	
TOGAGIAATTOGAGC	Aga	RIGODAY	30892 togostostestes 30678	20-1 1-0-1	Diasix (expect-100)	
TGGAGCTGATGGAGC	*		140008 transmitted to 100000	1,000.000	mile Intonposite, neliti	
TGGAGCAGCTGGAGC	Ser	AF028268	29993 topsersontess 20019	1,28.2-36.3	unel, Leu Zip p40,	
TOGAGCTGCTGGAGC	Š	AL009051	78023 trespetentment 78037	1981	metaxin	
TOGACCACCTOCACT	Tar	297200	112905 temporary temporary	10.5.24 10.5.24	HP-10, semaphorin F,G	
TGGAGTGCCTGGAGC	7	U95628	101031 teparterizes 101017	47b7	Akr enhancer BP, RAR	
TOGAGTAGATGGAGT	Arg	284495	2547 terasteraterast 2538	0.0.0	CFTC, TEK	
TOGAGCTGATGGAGT	*	274023	5255 terascicaterast 5241	Sp. 21.5	Po A B D alemand	
TOGAGTTTCTGGAGT	Phe	268275	7291 tggagtttctggagt 7277	4018.3	mens NAMAD	
TOGAGTGCCTGGAGT	Als	254072	21277 tgraftgottgraft 21291	4016.3	ork Acht Hens	
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TOGAGITACTOGAGI	ž	AC003951	27290 tggagttactggagt 27304	0	collaren	
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TUGACTIGATICAACC		L81613	2418 iggagitgalgrage 2404 ·	10	APC, bat2, p63	
TGGAGTGTATGGAGT	3	AC002122	43679 tggagtgtatggagt 43665	5p16.2	Met tRNA syntage	
TUGAGICCATUGAGT	er A	AC002380	34646 tggagtcentggagt 34632	6p16.2	N-WASP, enirma	
TUGAGCAACTGGAGC	Agn	AC002479	80443 tgrafcaactgrage 80457	5p15.2	NEU, alycoprotein C	
TOGAGCTGCTGGAGT	2 20	AC004892	125446 tggagetgetgagt 125431	6q31	CD22-B	
TOGAGTAGCTGGAGT	Ş	AC002393	3721 tegagtagutggagt 3735	. 9	Elycoprotein	
TGGAGTTGCTGGAGT	%	AC002326	114578 tggagttgetggagt 114564	9	G3P REGULON	
TGGAGTGCATGGAGT	Ala.	284490	20244 tggagtgcatggagt 20230		Alu, adrenargic receptor	

Table 2

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		640067	2080U iggageleitgrage 26788	6926-927	collagen, AT3, C1Ob
TARPACT CCITARAGI.	Ser	AC003090	22068 trraretectraret 22082	70.16	601
TGGAGTATATGGAGC	Пе	ACD04744	99740 tomorphis tomorphis	07.41	2
PGGAGTAGTAGGAGG		*********	POLICE CONTRACTOR AND	7p15-p21	TSH.R, RNABP
	9	AC004485	86356 iggagtagetggage 86370	7015-021	Hox 2.4. mII.11 Hofaton*)
TUGAGICITITGGAGT	ទី	AC004141	3130 temastetteraset 8144	7c91-n99	
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TOANGINACIOCACT	S.	AC000064	9170 tggagtaactgraat 9184	7421.22	C 4 9 D
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Table 3

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TGGAGCTGATGGAGC	Į	AC000061	79564 terserterstosen 10880	0.00	
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		AC000125	18750 tggagtttttggagt 18786	7431.3	p160
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TGGAGCATTTAGAGG	Ė	400004	Librar allegates and to con-	7435(TeRb)	properdin
	3	AC003109	4761 tggagcatttggage 4775	7936	CD2 HOX.2 &
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TOGAGTGCATGGAGT	Ala	ACOOSEO	99849 to		Transpired in the sale
	•		Topic illustifications of the contraction of the co	12924	Alu, IFNaR

Table 4

Probe .	Хав	Accession No.	Hit site	Locus	blastx (expect=100)
TOGAGTGCATGGAGT	Ala.	AC004217	88822 terartecaterart 88808	12024.1	Alu HPK
TTGGAGTTACTGGAGC	Ę.	AC002978	65893 teracttacterare 65907	12024	clathrin I.C. RPOR (non We)
TGGAGTTGTTGGAGT	Š	AC000403	91715 teractetteeset 91729	13	VHT. fnhihn B
TOGAGCGOTTGGAGC	ਰੰ	X97061	73621 tgracegiterare 73607	14032.33 Ge	14082.38 (JeD) polycyatic kidney
TGGAGTAGGTGGAGC	Ast	AC003024	15596 tggagtaggtggage 15552	16926	DkaP
TOGAGTTTCTGGAGC	Phe	ACROD 192	93356 tgragtttctggage 93370	16	HAND Tou.
TOGAGTTCATOGAGT	8	U91818	102406 tgragtcatgragt 102392	19	ICAM1, MIBP1
TOGACTOTATOCACT	Val	AC002289	10631 tggagtgtatggagt 10845	91	Alu
TOURDOOM				SHOW STATE	
TOGAGTTAATGGAGT	:	AC002519	81768 trgagttaatgragt 81754	16	Rho Notch
TOGAGCTGCTGGAGT	Š	U91826	84127 tegagoteoteract 84118	18011.2	NIPLINA II 9Rr(non WS)
TOGACTOAATGGAGT	Chu	ACOUSTOR	10952 termetrantment 10966	16012	
TOGAGCACTTOGAGC	결	AC002551	82245 treascactterare 82259	16012.1	envelone anderen P
TGGAGTCCCTGGAGC	F	AC002289	162 terartecterare 148	16012-018 1	_
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TOGACTCACTOGAGT	AH.	U96787		16p13.1	Tera, HLAs
			16374 tggagtenetggagt 16388		Notch, Pro-rich
			16599 tggagteactggagt 16613		phosphatese, ORFB
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Table 5

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TGGAGCCGCTGGAGC	Are	A CON1000		LOCUS	blastx (expect=100)
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TOGAGINTERCAGE		9990000	114346 iggagtaaatggagt 114860	11	bata-D-rincosidasa
	3	AC003967	52898 tggagtetetggage 52884	17	THE STATE OF THE
I GGAGCAGATGGAGC	Arg	AC003971	76277 terascosstscosco 2000		tie-1, SEA, Kho,
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1 CONCOUNT COACC	ş	AC004004	89010 teracraretersen 98000	96.96	
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		TOTAL PROPERTY.		-	CSFRb. IL11R(+ston codon)
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	į		31093 iggagitgatggage 31079	20912-18.2	Amenborin F GUG D 1477
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TGGAGTGCCTGGAGT	Ala	AF039907	20809 tree standard CO802	(190)17	OLL, HAIR ILTRIDONWS
TOGAGTGTCTGGAGT	Val		90887 188891 mg by 10 mg	.	IgV, Cyt.Oxidase
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Table 6

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. 15	blastx (expect=100)	Na/Ca exchanger	RNA polymerese	smaphorin F	Glveonrotain	MINICOLOGICAL DE	IcV.	Ir H. TCF-8. CETP	Alu, BCGF	TPO	semaphorin H. CD44	ERF	factor H	Alu, NF2	collagen al. Na channel	ADH, collagen	22911DiGeorgeclathrin heavy chain 2	IN CONTRACTOR	JeHv. PC binding	D150. II ARWSNWSF	FOFRb	collagen at	collagen al,
<i>2</i> 0	Locus	21q11.1	21911.1	21911.1	21911.1		21911.1	21q11.1	21411.1	21922.2	23	23	22q11.2-qter	22q11.2-qter	22011.2	22411.2	22q11DiGeorg	THE CONTRACT OF THE PROPERTY O	22011.2	22012.1	22q12,1-gter	22q12-gter	22q12-qter
25		itgengt 28789	tgragt 24914	tggagt 21707	lggagt 28150	THE STATE OF	Fagt 7218	tggage 93740	igrage 17567	teraft 48494	161692 iggagigagiggagi 151618	gaft 8489	tggagt 69311	t graft 36882	ctggagt 130727	grage 40691	tgrage 21074			tggage 64495	ttggage 114794	184gc 2661	ggagt 40839
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40	Xaa ,	I.V.s	Arg	d G	7		Ala	e E	Z Z	O.y	Olu	£	ਝੰ	Als	Ž	Pro	å		Sex	Asn	ŧ	Pro	Ser
45		GGAGTAAATGGAGT	TOGAGTAGGTGGAOT	TOOAGTOAGTGGAGT	GGAGTGTCTGGAGT		199AQTQCCTQQAQT	TGGAGCATTTGGAGC	199AGCCTCT0GAGC	rggagtgggtggagt	TGGAGTGAGTGGAGT	ragagetagtagt	NGQAGTGQGTGGAGT	regagtecategagt	TGGAGCCTCTGGAGT	TOGAGTCCCTGGAGC	TOGAGCATCTOGAGC		PGGAGCAGCTGGAGC	TGGAGCAACTGGAGC	TGGAGCTAGTGGAGC	TOGAGCCCTTGGAGC	TGGAGCTCTTGGAGT
50	Probe	TOGAGI	TTOGAG	TOGAGT	TOGAGT		TOGAGT	TOGAGO	TOGAGO	TOGACT	TGGAGT	TOGAGO	TOGAGT	TOGAGI	TGGAGC	TOGAGT	TOGAGC		TGGAGC	TGGAGC	TGGAGC	TGGAGC	TGGAGC

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Table 7

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Probe	Xaa	Accession No.	Hit site	Locus	blastx (expect=100)
TOGAGCCATTGGAGT	His	Z81808	12676 tggegccattggegt 12561	22q12-qter	MYF:5, p53, INK4a
PREACTORACTORACT	Cla	ALBONG317	85322 tegangear (repair 85336	22012 3-1112	22012 3-14 2 CM-CSFRD M. BREPOR, or
TTGGAGTGAGTGGAGT	g	U62817	77740 tegagteagtegest 77726	22413	latrophilin-related
TOGAGTGCATGGAGT	Ala	Z980 IS	31082 tgraftfcatgraft 31068	22418	Alu, DECEMBE, ADJC-NTP
TOGAGTTOTTGGAGT	Š	AC002422	19151 tggagttgttggagt 19137	×	comP PDate
TOGAGTOTCTGGAGT	Vel	273418	31830 trgartetetrage 31816	×	WNT-8D, MI-2
TGGAGTCTTTGGAGT	197	283848	114972 tggagtettiggagt 114955	×	reverse transcriptase
TGGAGTCTCTGGAGT	Lea	901682	7749 tggagtetetggagt 7785	*	Selenoprotein
TOGAGCAACTGGAGT	Asn	AC002420	70704 tgrageactgragt 70890	×	homeoprotein. OBR(aton)
TOGAGCATOTGGAGT	Met	01.27.77	5702 teracateterati 5859	*	Terb, Differen
TOGAGITTCCTGGAGC	Ser	283131	4904 tggagttectggage 4890	×	VPS41 homolog
TOGAGTGGCTGGAGC.	ð	AC004388	239975 terartereterare 239989	×	GAP mLIFR(ston)
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TGGAGTCTATGGAGC	Zer.		9984 tggagtetatggage 9948	×	complement C8, C7
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TOGAGCTGTTGGAGC	Ç,	L44140	112657 tggagetgttggage 112671	×	rab GDI alpha, BDGF
TOGAGCTCATGGAGC	Ser	AC004383	144906 tggagctcatggage 144892	×	RTass, transposon
TOGAGTAAATOGAGC	ŗ.	269732	31681 tggagtanatggage 31695	Xp11	OPR, acrosin
TOGAGTTCGTGGAGC	Ser	292545	88703 tggagticgtggage 88717	Xp11	PMK1
TGGAGCTTCTGGAGC	Phe	AL008709	46089 tggagetetggage 48075	Xp11.23-Xp1	Xp11.23-Xp11.4rMHC class 1a, HLA-C
TOGAGITITCTGGAGT	Phe	U96409	116332 tgregtticigregt 116346	Xp22	myosin H
TOGAGTTGCTTGCAGT	Č	ACHORITOR	89544 terretecterant 89530	Xn22	

.

Table 8

Probe	Хав	Xaa Accession No. Hit site	Hit site	Locus	blastx (expect=100)
TTGGAGTCACTGGAGT	H	AL021706	11982 tggagteactggagt 11968	Xq21.1-21.89	Xq21,1-21,83 dopamine receptor
TOGAGCTGGTGGAGT	Ę	AC000113	119188 tggagotggtgragt 119202	Xq23	DNA repair protein, MHC
TGGAGCAAGTGGAGT	Š	AF007262	96212 tggsgcaagtggagt 98226	Xq28	RNA polymerase
TOGAGCTGCTGGAGT	Š	U82671	35792 tggagetgetggagt 35806	Xq28	XTCF-8c
TOGAGTCAGTGGAGC	9	AP011889	144485 tggagtengtggage 144451	Xq28	CHRHR, Werner Synd.
TGGAGCTAATGGAGC	i	AF030876	107409 tgragetaatgrage 107895	Xq28	gp41, clk3
			出ているとうが、100mmに対象を対することに対している。 100mmであるというでは、100mmでは	and the state of	
	2				
TOGAGITICTGGAGI	Pbe	AC002531	106695 tggagttctggagt 106712	> -	Alu, hpk
TGGAGCAGTTGGAGC	2961	AC004474	124745 tgagcagtiggage 124731	> -	EGFR, Smad6
TOGAGTTTGTGGAGT	I.eu	1126428	12899 tesestattataess 12991	מיט זמ	DBT D(manages)
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והפשפרשערו הפשפו	2	02/960	e 1972 terancellare 61658	Bouge DNA	envelope mil. 11 Kionnasite)
TGGAGTCCCTGGAGC	£	0.35323	22244 tgragtocotggage 22230	MHC cless II	CFTC, United
TOGAGCAGATGGAGC	Arg	AC002482	14276 tgragcagatgrago 14290	RG208003	I-309, TeR, IL9R(nonWS)
TOGAGCTCTTGGAGC	Ser	U34879	24914 tggagetettggage 24928	EDH1782	Large tegument protein
					common Bloppsit, non WS)
TOGAGCCTTTGGAGC	នឹ	216028	6359 tggageettiggage 6373	Bat2	bat2,mucin,
GM-CSFRb(onnosite_ston)					

Redundant clones are shadowed. White and underlined letters indicate hits and pseudo-hits, respectively.

[0140] Four clones out of these 28 clones (AC002303, AC003112, AL008637, and AC004004) hit several known hemopoietin receptors, however, AC004004 was excluded as it has a stop codon downstream three amino acids of the

Trp-Ser-Xaa-Trp-Ser motif. Among the three remaining clones, AL008637 was thought to be a known receptor, GM-CSF receptor β. AC002303 is the BAC clone CIT987-SKA-67085 derived from the 16p12 region of human chromosome no. 16 registered by TIGR group on June 19, 1997 and comprises the full-length of 131530 base pairs (Lamerdin, J.E., et al., GenBank Report on AC003112, 1997).

[0141] As shown in Fig. 1, a BlastX search (query: 180 nucleotides of 40861-41040 including tggagtgaatggagt (40952-40966), the only probe sequence within the AC002303) revealed that numerous hemopoietin receptors starting with the TPO receptor and leptin receptor show an evident homology, however, there were no known, database-registered hemopoietin receptors that completely matched the query sequence. Also, a BlastX scanning was done under the above conditions, by excising a sequential 180-residue nucleotide sequence in both the 5' and 3' directions, centering on the 180-residue nucleotide sequence mentioned above, and when this was used as a query, two sequences having a homology to known hemopoietin receptors were found in the regions 39181-39360 and 42301-42480, and were thought to be other exons of the same gene (Fig. 2).

[0142] A Pro-rich motif PAPPF was conserved in the 39181-39360 site, and a Box 1 motif in the 42301-42480 site. The 3' side exon adjacent to the exon containing the Trp-Ser-Xaa-Trp-Ser motif has a transmembrane domain, and this domain has a low homology with other hemopoletin receptors, and was not detected by the BlastX scan. These results suggested the possibility of a novel hemopoletin receptor gene existing in the above-described BAC clone CIT987-SKA-670B5.

Example 2: Search for NR8 expressing tissues using RT-PCR

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[0143] Pseudogenes have been reported to exist in several hemopoietin receptors (Kermouni, A. et al., Genomics, 1995, 29 (2) 371-382; Fukunaga, R. and Nagata, S., Eur. J. Biochem., 1994, 220, 881-891). To verify that NR8 is not a pseudogene, and with the objective of identifying NR8 expressing tissues, transcripts of the NR8 gene were searched by RT-PCR method.

In the AC002303 sequence of the above-described BAC clone, several exon regions widely conserved at the amino acid translation level in known cytokine receptors were surmised, and on the sequence of the surmised exon region, the following primers were synthesized. (See Fig. 5 for the location of each primer.)

NR8-SN1; 5'- CCG GCT CCC CCT TTC AAC GTG ACT GTG ACC -3' (SEQ ID NO: 9)

NR8-SN2; 5'- GGC AAG CTT CAG TAT GAG CTG CAG TAC AGG -3' (SEQ ID NO: 10)

NR8-AS1; 5'- ACC CTC TGA CTG GGT CTG AAA GAT GAC CGG -3' (SEQ ID NO: 11)

NR8-AS2; 5'- CAT GGG CCC TGC CCG CAC CTG CAG CTC ATA -3' (SEQ ID NO: 12)

[0145] Using the Human Fetal Multiple Tissue cDNA Panel (Clontech #K1425-1) as the template, RT-PCR was attempted using combinations of the above primers. Advantage cDNA Polymerase Mix (Clontech #8417-1) was used for the PCR, which was conducted under the conditions below using the Perkin Elmer Gene Amp PCR System 2400 Thermalcycler.

[0146] Namely, the PCR conditions were, 94°C for 4 min, 5 cycles of "94°C for 20 sec, 72°C for 3 min," 5 cycles of "94°C for 20 sec, 70°C for 3 min," 28 cycles of "94°C for 20 sec, 68°C for 3 min," 72°C for 4 min, and completed at 4°C.

[0147] From the primer locations shown in Fig. 5, amplifications of bands sized 330 bp, 258 bp, 234 bp, and 162 bp can be expected from the combinations of SN1/AS1, SN1/AS2, SN2/AS1, and SN2/AS2. When evaluated using human fetal liver, brain, and skeletal muscle cDNA as the template, clear bands having the anticipated sizes were obtained in the fetal liver only with the respective primer combinations (Fig. 3).

[0148] An amplification was not seen at all for fetal brain cDNA, and a band of about 650 bp and a broad band of 400 to 500 bp were observed for fetal skeletal muscle cDNA. However, since the band sizes for skeletal muscle cDNA remained constant even when different combinations of primers were used, it is thought that these bands were non-specific amplifications due to some reason.

[0149] The obtained PCR product was subcloned to pGEM-T Easy vector (Promega #A1360), and the nucleotide sequence was determined. The recombination of PCR products to the pGEM-T Easy vector was done by T4 DNA Ligase (Promega #A1360) reacted at 4°C for 12 hr. The genetic recombinant between the PCR product and pGEM-T Easy vector was obtained by transforming *E.coli* strain DH5 α (Toyobo #DNA-903).

[0150] For the selection of the genetic recombinant, Insert Check Ready (Toyobo #PIK-101) was used. The dRhod-amine. Terminator Cycle Sequencing Kit (ABI/Perkin Elmer #4303141) was used for determining the nucleotide sequence, and analysis was done using the ABI PRISM 377 DNA Sequencer. As a result of determining the nucleotide sequences of all inserts of the 10 independent clones of genetic recombinants, all clones were found to comprise a single nucleotide sequence. These obtained sequences were verified to be partial nucleotide sequences of NR8.

Example 3: Full-I ngth cDNA cloning by the 5' and 3'-RACE methods

[0151] Using the thus-obtained fetal liver-derived cDNA, 5' and 3'-RACE methods were conducted to obtain full-length cDNA (Fig. 4).

3-1) 5'-RACE method

[0152] 5'-RACE PCR was performed using the above-mentioned NR8-AS1 primer for primary PCR, and NR8-AS2 primer for secondary PCR. Human Fetal Liver Marathon-Ready cDNA Library (Clontech #7403-1) was used as the template and Advantage cDNA Polymerase Mix for the PCR experiment. As a result of PCR under the following conditions using the Perkin Elmer Gene Amp PCR System 2400 Thermalcycler, two types of PCR products were obtained, which have different sizes through selective splicing.

[0153] Primary PCR conditions were 94°C for 4 min, 5 cycles of "94°C for 20 sec, 72°C for 4 min," 5 cycles of "94°C for 20 sec, 70°C for 4 min," 28 cycles of "94°C for 20 sec, 68°C for 4 min," 72°C for 4 min, and completed at 4°C.

[0154] Secondary PCR conditions were 94°C for 4 min, 5 cycles of "94°C for 20 sec, 70°C for 3 min 30 sec," 28 cycles of "94°C for 20 sec, 68°C for 3 min 30 sec," 72°C for 4 min, and completed at 4°C.

[0155] Both types of PCR products obtained were subcloned to pGEM-T Easy vector as mentioned earlier, and the nucleotide sequences of all inserts were determined for the 16 independent clones of genetic transformants. As before, the dRhodamine Terminator Cycle Sequencing Kit was used for determining the nucleotide sequence, and analysis was done using the ABI PRISM 377 DNA Sequencer. As a result, the clones can be divided into two groups, one having 14 clones, and the other having 2 clones, by the length of the base pairs and the differences in sequence (though described later, the differences lie in the products due to selective splicing, and the group of 14 independent clones comprises the sequence corresponding to exon 5 in the genomic sequence, and the remaining group of two independent clones does not have this sequence).

3-2) 3'-RACE method

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[0156] 3'-RACE PCR was performed using the above-mentioned NR8-SN1 primer for primary PCR, and NR8-SN2 primer for secondary PCR. Human Fetal Liver Marathon-Ready cDNA Library was used as the template similar to 5'-RACE PCR, and Advantage cDNA Polymerase Mix for the PCR experiment. As a result of conducting PCR under the conditions shown in 3-1), a single band PCR product was obtained.

[0157] The obtained PCR product was subcloned to pGEM-T Easy vector as above, and the nucleotide sequences of all inserts of the 12 independent clones of genetic recombinants were determined. As before, the dRhodamine Terminator Cycle Sequencing Kit was used for determining the nucleotide sequence, and the sequences determined were analyzed using the ABI PRISM 377 DNA Sequencer. As a result, all 12 independent clones showed a single nucleotide sequence.

[0158] As a result of analyzing the nucleotide sequence of the fragments (approximately 1.1 kb and 1.2 kb) amplified by 5'-RACE and 3'-RACE, respectively, it was conceived that the approximately 260 bp of each fragment overlap and extend to the 5' side and 3' side, and contain almost the full-length of NR8 mRNA. These were joined to make a full-length cDNA (NR8\alpha) (Fig. 5 and Fig. 6). The plasmid containing the NR8\alpha cDNA (SEQ ID NO: 2) was named pGEM-NR8\alpha, and *E.coli* containing the plasmid has been internationally deposited at the National Institute of Bio-Science and Human-Technology, Agency of Industrial Science and Technology (1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan) under the accession number FERM BP-6543 since October 9, 1998 according to the Budapest Treaty.

[0159] As shown in Fig. 5 and Fig. 6, in the ORF of NR8α cDNA, the Met starting from nucleotide no. 441 is thought to be the start codon due to the presence of an inframe stop codon 39 bp upstream, and completes with two stop codons starting from nucleotide no. 1524. It has the features of, from the N terminus in order, a typical secretion signal sequence, a domain thought to be the ligand binding site containing a Cys residue conserved in other hemopoietic receptor members, a Pro-rich motif, Trp-Ser-Xaa-Trp-Ser motif, a transmembrane domain, a Box 1 motif thought to be involved in signal transduction, and such features of hemopoietin receptors. From the above results, the NR8 gene was thought to encode a novel hemopoietin receptor.

[0160] Analysis of fragments amplified by the RACE method suggested the presence of a splice variant. As a result of nucleotide sequence analysis, this variant was revealed to be lacking approximately 150 bp including the above-described Pro-rich motif of NR8 α . Moreover, as a result of comparing AC002303 sequence with NR8 α , and carrying out analogy of exons/introns (Table 9), the above-described variant was thought to be deficient of the 5th exon due to selective splicing.

Table 9

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Characteristics	inframe stop codon	start codon, signal peptide	conserved Cys residue	conserved Cys residue, N-glycosylation site	Pro-rich motif (PAPPF), N-glycosylation site	grWSEWSdp motif	transmembrane domain	Box1 (IWAVPSP)	connects to exon 10, Box2-like sequence (PSTLEVYSCH), nontypical exon/intron boundary	double stop codons, Box2-like sequence (PSTLEVYSCH, PAELVESDG), polyA	double stop codons, polyA	
# in NR8	:1-424	: 425-489	: 490-592	: 593-792	: 793-947	: 948-1125	: 1126-1225	: 1226-1307	: 1308-1405*	: 1308-2465**	: 1406-1934*	5+6+7+8+9a+1(
# in AC002303	4	26334-26398	30625-30727	33766-33965	39240-39394	40820-40997	41455-41554	42285-42366	44812-44909	44812-45922<	45441-45922<	NR8 a*: exons 1+2+3+4+5+6+7+8+9a+10
Exon	1	,	m	4	8	9	7	00	. 9a	96	10	NR8 a*

(two alternative reading frames for soluble-type and transmembrane(-signal)-type)
NR8 +**: exons 1+2+3+4+5+6+7+8+9b

NR8 \(\beta\): exons 1+2+3+4+6+7+8+9a+10

[0161] This variant (NR8 β) can encode a soluble receptor in the truncated form by the joining of the 6^{th} exon directly to the 4^{th} exon and causing a frame shift. The boundary between the exons and the introns takes a consensus

sequence in most cases, but the boundary betw en the 9th exon (Exon 9a) and the 9th intron is the only boundary that takes a different sequence from the consensus sequence (nag/gtgagt, etc.), being acc/acggag. The plasmid comprising NR8β cDNA (SEQ ID NO: 4) was named pGEM-NR8β, and *E.coli* comprising the plasmid has been internationally deposited at the National Institute of BioScience and Human-Technology, Agency of Industrial Science and Technology (1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan) under the accession number FERM BP-6544 since October 9, 1998 according to the Budapest Treaty.

Example 4: Northern blotting

[0162] In order to analyze the distribution and mode of NR8 gene expression in each human organ and human cancer cell lines, Northern blot analysis was done using the cDNA encoding the full-length NR8α protein prepared based on all the cDNA fragments obtained in Example 3 as a probe. The probe was prepared using Mega Prime Kit (Amersham, cat#RPN1607) by radiolabeling it with [α-32P] dCTP (Amersham, cat#AA0005).

[0163] As Northern blots, Human Multiple Tissue Northern (MTN) Blot (Clontech #7760-1), Human MTN Blot IV (Clontech #7766-1), and Human Cancer Cell Line MTN Blot (Clontech #7757-1) were used. Express Hyb Hybridization Solution (Clontech #8015-2) was used for hybridization.

[0164] Hybridization conditions were: a prehybridization at 68°C for 30 min, followed by hybridization at 68°C for 14 hr. After washing under the following conditions, the blots were exposed to Imaging Plate (FUJI#BAS-III), and the gene expression of NR8 mRNA was detected by the Image Analyzer (FUJIX, BAS-2000 II). Washing conditions were: (1) 1x SSC/0.1% SDS, at room temperature for 5 min; (2) 1x SSC/0.1% SDS, at 50°C 30 min; and (3) 0.1x SSC/0.1% SDS, at 50°C 30 min.

[0165] Fig. 12 shows the results of Northern blot analysis of NR8 expression in each organ. A total of three different-sized mRNA, one 5kb-sized and two 3 to 4kb sized, were detected in human adult lung, spleen, thymus, skeletal muscle, pancreas, small intestines, peripheral leucocytes, and uterus. A similar examination of various cell lines including hemopoietic cell lines showed similar sized bands in two cell lines, the promyeloid leukemic cell line HL60 and Burkett's lymphoma-derived Raji.

Example 5: Plaque screening

Northern blot analysis of NR8 gene expression detected at least three types of specific mRNA bands with different sizes in each human organ and in each human cancer cell line for which NR8 gene expression was seen. However, the inventors had succeeded in isolating only two types of selective splicing variants, namely NR8α and NR8β genes, in the above-described Examples. Therefore, the inventors performed plaque screening with the objective of isolating the gene of the third selective splicing variant. Human Lymph Node (Clontech, cat#HL5000a) that showed a strong NR8 gene expression in the above-mentioned Northern analysis results, was used as the cDNA library. The probe used was NR8α cDNA fragment, which was radio-labeled by [α-³²P] dCTP (Amersham, cat#AA0005) using the Mega Prime Kit (Amersham, cat#RPN1607). Approximately 7.2 x 10⁵ plagues of Human Lymph Node cDNA Library were blotted onto a Hybond N (+) (Amersham, cat#RPN303B) charged nylon membrane to conduct primary screening. Rapid Hybridization Buffer (Amersham, cat#RPN1636) was used for the hybridization. Hybridization conditions were: a prehybridization at 65°C for 1 hr, followed by hybridization at 65°C for 14 hr. After washing under the conditions, (1) 1x SSC/0.1% SDS, at room temperature for 15 min; (2) 1x SSC/0.1% SDS, at 58°C 30 min; and (3) 0.1x SSC/0.1% SDS, at 58°C 30 min, the membrane was exposed to an X-ray film (Kodak, cat#165-1512) to detect NR8 positive plaques. [0167] As a result, positive or pseudo-positive 16 independent clones were obtained. A similar secondary screen-

ing was done for the 16 clones obtained from the primary screening to successfully isolate plaques of NR8 positive 15 independent clones. The inserts of these 15 clones were amplified by PCR through a pair of primers located in both ends of the \(\lambda\)gt10 vector cloning site. Advantage cDNA polymerase Mix (Clontech #8417-1) was used for the PCR reaction conducted using the Perkin Elmer Gene Amp PCR System 2400 Thermalcycler, under the following experiment conditions. Namely, 94°C for 4 min, 5 cycles of "94°C for 20 sec, 70°C for 4 min," 30 cycles of "94°C for 20 sec, 68°C for 4 min, and completed at 4°C.

[0168] Similar to above, the obtained PCR products were subcloned to pGEM-T Easy vector, and the nucleotide sequence of the inserts were determined using the BigDye Terminator Cycle Sequencing SF Ready Reaction Kit (ABI/Perkin Elmer#4303150), and analyzed by the ABI PRISM 377 DNA Sequencer. As a result, among the 15 clones obtained, at least two clones showed an insertion of 177 amino acids flanking the NR8α C terminus, and since this portion derives from the 9th intron of the NR8 gene and is removed by splicing in NR8α, this 3rd selective splicing variant was named NR8γ. The plasmid containing the NR8γ cDNA (SEQ ID NO: 8) was named pGEM-NR8γ, and *E.coli* containing the plasmid has been internationally deposited at the National Institute of BioScience and Human-Technology, Agency of Industrial Science and Technology (1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan) under the accession number FERM BP-6545 since October 9, 1998 according to the Budapest Treaty.

[0169] Among the 15 clones obtained here, four clones other than the two mentioned above were further selected, and their nucleotide sequences were analyzed. As a result, among the six clones selected, two clones had the NR8 β nucleotide sequence, and all the remaining four clones had the NR8 γ nucleotide sequence. Therefore, the six clones for which the nucleotide sequence was analyzed did not contain the NR8 α sequence. The NR8 γ cDNA clones for which the nucleotide sequences were determined included those having 3'-UTR (3UTR-2) in which a poly-A tail is added to the site elongated 483 bp from the 3'-UTR of NR8 α obtained by the 3'-RACE method (3UTR-1), and those having 3'-UTR (3UTR-3) in which a poly-A tail is added to the site elongated 2397 bp from the 3'-UTR of NR8 α . On the other hand, the two clones of NR8 β for which the nucleotide sequence was decided above, both contained the nucleotide sequence of 3UTR-3. In Table 10 below, the 3' end non-translation region sequences contained in the cDNA clones thus far obtained are summarized. Also, the nucleotide sequences of 3UTR-1, 3UTR-2, and 3UTR-3 following the translation stop codon of NR8 γ cDNA sequence are shown in SEQ ID NO: 23, SEQ ID NO: 24, and SEQ ID NO: 25, respectively.

[0170] Moreover, the nucleotide sequences of 3UTR-B1 and 3UTR-B3 following the translation stop codon of NR8β cDNA sequence are shown in SEQ ID NO: 26 and SEQ ID NO: 27, respectively.

Table 10

NR8 cDNA clone	3'-UTR sequence
NR8α	3UTR-1
NR8ß	3UTR-B1, 3UTR-B3
NR8y	3UTR-1, 3UTR-2, 3UTR-3

[0171] The nucleotide sequences thus obtained revealed that the gene transcripts of NR8 can encode various different sizes not only due to the differences in selective splicing, but also due to the length of the 3' end non-translation region sequence. This may adequately explain the presence of various-sized transcripts detected by Northern blot analysis.

Example 6: Ligand screening

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6-1) Construction of NR8 chimeric receptor

A screening system was constructed for searching a ligand that can specifically bind to NR8, namely, a novel hemopoietin. First, the cDNA sequence encoding the extracellular region of NR8α (the amino acid sequence of SEQ ID NO: 1; from the 1st Met to the 228th Glu) was amplified by PCR, and this DNA fragment was bound to DNA fragments encoding the transmembrane region and the intracellular region of a known hemopoietin receptor to prepare a fusion sequence encoding a chimeric receptor. As described above, there were several candidates for the partner, the known hemopoietin receptor, and among them, the human TPO receptor (Human MPL-P) was selected. Namely, after amplifying the DNA sequence encoding the intracellular region that includes the transmembrane region of the human TPO receptor by PCR, this sequence was bound to the cDNA sequence encoding the extracellular region of NR8α in frame, and inserted into a plasmid vector expressible in mammalian cells. The expression vector constructed was named pEF-NR8/TPO-R. A schematic diagram of the structure of the constructed NR8/TPO-R chimeric receptor is shown in Fig. 14, and the nucleotide sequence of the chimeric receptor and the expressible amino acid sequence encoded by it are shown in SEQ ID NOs: 13 and 14, respectively. Together with an expression vector pSV2bsr (Kaken Pharmaceutical Co., Ltd.) containing Blastcidin S resistant gene, the NR8/TPO-R chimeric receptor-expressing vector was introduced into the growth factor-dependent cell line Ba/F3, and forcedly expressed. Gene-introduced cells were selected by culturing with 8 µg/ml of Blastcidin S hydrochloride (Kaken Pharmaceutical Co., Ltd.) and IL-3. By transferring the obtained chimeric receptor-introduced cells to an IL-3-free medium, adding a material expected to contain a target ligand, and culturing, it is possible to conduct a screening that uses the fact that survival/proliferation will be possible only when a ligand that specifically binds to NR8 is present.

6-2) Preparation of NR8/IgG1-Fc soluble fusion protein.

[0173] NR8/lgG1-Fc soluble fusion protein was prepared to be used for searching cell membrane-bound ligands, or the detection of soluble ligands through BIAcore (Pharmacia) and West-western blotting. A fusion sequence encoding the soluble fusion protein was prepared by binding a DNA fragment encoding the extracellular region of NR8α (amino acid sequence; from the 1st Met to the 228th Glu) prepared in 5-1) with the DNA fragment encoding the Fc

region of human immunoglobulin IgG1 in frame. A schematic diagram of the structure of the soluble fusion protein encoding the NR8/IgG1-Fc is shown in Fig. 14, and the nucleotide sequence and the expressible amino acid sequence encoded by it in SEQ ID NOs: 15 and 16, respectively. This fusion gene fragment was inserted into a plasmid vector expressible in mammalian cells, and the constructed expression vector was named pEF-NR8/IgG1-Fc. If this pEF-NR8/IgG1-Fc is forcedly expressed in mammalian cells, and after selecting stable gene-introduced cells, the recombinant protein secreted into the culture supernatant can be purified by immunoprecipitation using anti-human IgG1-Fc antibody, or by affinity columns, etc.

6-3) Construction of an expression system of NR8ß and purification of recombinant NR8ß protein

[0174] The recombinant NR8 β protein was prepared to be used for searching cell membrane-bound ligands, or the detection of soluble ligands using BlAcore (Pharmacia) or West-western-blotting. Using the amino acid coding sequence of NR8 β cDNA, the stop codon was replaced by point mutation to a nucleotide sequence encoding an arbitrary amino acid residue, and then, was bound to the nucleotide sequence encoding the FLAG peptide in frame. This bound fragment was inserted into a plasmid vector expressible within mammalian cells, and the constructed expression vector was named pEF-BOS/NR8 β FLAG. Fig. 14 shows a schematic diagram of the structure of the insert NR8 β FLAG within the constructed expression vector. Moreover, the nucleotide sequence of NR8 β FLAG and the expressible amino acid sequence encoded by it are shown in SEQ ID NOs: 17 and 18, respectively. If this pEF-BOS/NR8 β FLAG is forcedly expressed in mammalian cells, and after selecting stable gene-introduced cells, the recombinant protein secreted into the culture supernatant can be immunoprecipitated using anti-FLAG peptide antibody, or may be purified by affinity columns, etc.

Example 7: Isolation of mouse NR8 (mNR8) gene

7-1) The mouse homologous gene using human NR8 primers

[0175] Xenogeneic cross PCR cloning was isolated using the oligonucleotide primers, NR8-SN1 and NR8-SN2 (SEQ ID NOs: 9 and 10) at the sense side (downstream direction) and NR8-AS1 and NR8-AS2 (SEQ ID NOs: 11 and 12) at the antisense side (upstream direction), which were used for isolating full-length cDNA of human NR8. By combining the above-mentioned human NR8 primers, four types of primer sets can be constructed. Namely, using the combinations of "NR8-SN1 vs. NR8-AS1," "NR8-SN1 vs. NR8-AS2," "NR8-SN2 vs. NR8-AS1," and "NR8-SN2 vs. NR8-AS2," and a mouse brain cDNA library (Clontech #7450-1) and a mouse testis cDNA library (Clontech #7455-1) as templates, amplification of cross PCR products was expected. Advantage cDNA Polymerase Mix (Clontech #8417-1) was used for the PCR that was conducted under the conditions below using the Perkin Elmer Gene Amp PCR System 2400 Thermalcycler to amplify partial nucleotide sequence that could encode a mouse homologous gene of this receptor.

[0176] Namely, the cross PCR conditions were 94°C for 4 min, 5 cycles of "94°C for 20 sec, 72°C for 1 min," 5 cycles of "94°C for 20 sec,70°C for 1 min," 28 cycles of "94°C for 20 sec,68°C for 1 min," 72°C for 4 min, and completed at 4°C.

[0177] As a result, as shown in Fig. 15, an amplification of the cross PCR product was seen when any primer set was used. Also, a much clearer amplification product can be obtained when mouse brain cDNA was used as the template than when mouse testis cDNA was used.

7-2) Determination of the partial nucleotide sequence of the mouse homologous gene corresponding to NR8

Fig. 10178] Among the amplification products obtained in 7-1), mouse brain cDNA-derived product was subcloned to pGEM-T Easy vector (Promega #A1360), and the nucleotide sequence was determined. Namely, the PCR product was recombined into pGEM-T Easy vector by using T4 DNA ligase (Promega #A1360) at 4°C for 12 hr, and the resulting product was transfected into *E.coli* strain DH5α (Toyobo #DNA-903) to obtain the genetic recombinants of the PCR product and pGEM-T Easy vector. For the selection of genetic recombinant, Insert Check Ready Blue (Toyobo #PIK-201) was used. The nucleotide sequence was determined by using the BigDye Terminator Cycle Sequencing Ready Reaction Kit (ABI/Perkin Elmer #4303154), and sequence analysis was done by the ABI PRISM 377 DNA Sequencer. As a result of determining the nucleotide sequence of all inserts of eight independent clones of genetic recombinants, nucleotide sequences derived from the same transcript were obtained, and they were verified to be partial nucleotide sequences of mNR8. The obtained partial nucleotide sequence is shown in SEQ ID NO: 28.

7-3) Design of oligonucleotide primers specific to the mouse NR8 gene

[0179] Based on the partial nucleotide sequence of mNR8 obtained in 7-2), oligonucleotide primers specific to the

mouse NR8 were designed. As shown in the sequence given below, mNR8-SN3 was synthesized in the sense side (downstream direction), and, mNR8-AS3 was synthesized in the antisense side (upstream direction). ABI's 394 DNA/RNA Synthesizer was used for primer synthesis, which was done under 5'-end trityl residue addition conditions. After that, the complete length of the synthesized product was purified by using an OPC column (ABI #400771). These primers contributed towards the 5'-RACE method and the 3'-RACE method described later on.

mNR8-SN3; 5'- TCC AGG CGC TCA GAT TAC GAA GAC CCT GCC -3' (SEQ ID NO: 29) mNR8-AS3; 5'- ACT CCA GGT CCC CTG GTA GGA GGA GCC AGG -3' (SEQ ID NO: 30)

7-4) Cloning of cDNA corresponding to N terminus by the 5'-RACE method

[0180] To isolate full-length cDNA of mNR8, 5'-RACE PCR was performed using the NR8-AS2 primer (SEQ ID NO: 12) for the primary PCR, and the above-mentioned mNR8-AS3 primer (SEQ ID NO: 30) for secondary PCR. Mouse Brain Marathon-Ready cDNA Library (Clontech #7450-1) was used as the template, and Advantage cDNA Polymerase Mix for PCR experiment. As a result of conducting PCR under the following conditions using the Perkin Elmer Gene Amp PCR System 2400 Thermalcycler, PCR products of two different sizes were obtained.

[0181] Primary PCR conditions were 94°C for 4 min, 5 cycles of "94°C for 20 sec, 72°C for 100 sec," 5 cycles of "94°C for 20 sec,70°C for 100 sec," 28 cycles of "94°C for 20 sec,68°C for 100 sec," 72°C for 3 min, and completed at 4°C.

[0182] Secondary PCR conditions were 94°C for 4 min, 5 cycles of "94°C for 20 sec, 70°C for 100 sec," 25 cycles of "94°C for 20 sec,68°C for 100 sec," 72°C for 3 min, and completed at 4°C.

[0183] Both types of PCR products obtained were subcloned to pGEM-T Easy vector as described above, and the nucleotide sequences were determined. Namely, the PCR products were recombined into the pGEM-T Easy vector with T4 DNA ligase at 4°C for 12 hr, and the resulting product was transfected into *E.coli* strain DH5α to obtain the genetic recombinant between the PCR product and pGEM-T Easy vector. Also, as mentioned earlier, Insert Check Ready Blue was used for the selection of the genetic recombinant. For the determination of the nucleotide sequence, the BigDye Terminator Cycle Sequencing Ready Reaction Kit was used, and the nucleotide sequence was analyzed by the ABI PRISM 377 DNA Sequencer. The result of determining the nucleotide sequences of all inserts of eight independent clones of genetic recombinants suggests that they could be divided into two groups of four clones each by the base pair length and differences in the sequence. This difference of the products was caused by selective splicing, and both of the obtained sequences were verified to contain the sequence of full-length mNR8 cDNA clone corresponding to the N terminal sequence. The cDNA clone comprising the long ORF containing the exon encoding the Pro-rich region was named mNR8γ, and the cDNA clone encoding the short ORF that does not have the Pro-rich region was named mNR8β, respectively.

7-5) Cloning of cDNA corresponding to C terminus using the 3'-RACE method

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[0184] To isolate full-length cDNA of mNR8, 3'-RACE PCR was performed using the NR8-SN1 primer (SEQ ID NO: 9) for the primary PCR, and the mNR8-SN3 primer (SEQ ID NO: 29) for secondary PCR. Mouse Brain Marathon-Ready cDNA Library was used as the template, and Advantage cDNA Polymerase Mix for PCR experiment. As a result of conducting PCR under the above-mentioned conditions using the Perkin Elmer Gene Amp PCR System 2400 Thermalcycler, a PCR product of a single size was obtained. The PCR product obtained was subcloned to pGEM-T Easy vector as before according to 7-2), and the nucleotide sequence was determined. As a result of determining the nucleotide sequences of all inserts of four independent clones of genetic recombinants, it was found to contain the sequence of full-length mNR8 cDNA corresponding to the C terminal sequence. By combining the resulting nucleotide sequence determined through this 3'-RACE PCR, and the nucleotide sequence of 5'-RACE PCR products determined in 7-4), the complete nucleotide sequences of the full-length of mNR8γ and mNR8β cDNA were finally determined. The determined mNR8γ cDNA nucleotide sequence and the amino acid sequence encoded by it are shown in SEQ ID NOs: 22 and 21, respectively. The determined mNR8β cDNA nucleotide sequence and the amino acid sequence encoded by it are shown in SEQ ID NOs: 20 and 19, respectively.

[0185] When the human and mouse NR8 amino acid sequences were compared, a high homology of 98.9% was seen for NRθγ, and the homology was 97.2% even for NRθβ. This result strongly suggests the possibility that the same receptor gene has a vital functional responsibility that exceeds species. Fig. 16 shows a comparison between human and mouse NR8β amino acid sequences. Fig. 17 shows a comparison between human and mouse NR8γ amino acid sequences.

[0186] Both the full-length cDNAs of mNR8γ and mNR8β finally isolated were able to encode the transmembrane receptor protein comprising 538 amino acids, and the soluble receptor-like protein comprising 144 amino acids, respectively, through a selective splicing similar to human NR8. The structure below shows the characteristics of mNR8γ. First,

it is presumed that from amino acid no. 1 Met to amino acid no. 19 Gly is a typical secretion signal sequence. Here, since an inframe stop codon exists in the minus 13 position from the 1st Met, this Met residue is presumed to be the translation start codon. Next, from the 25th Cys to the 35th Cys residue is a typical ligand binding site sequence, and the 65th and 109th Cys residues also show the repetitive Cys residue structure conserved in other hemopoietin receptors as well. Next, the Pro-rich region is conserved by the Pro residues repeating at the 120th, 122rd and 123rd positions. From the 214th Trp to 218th Ser residue is a typical WSXWS-Box (WS motif). Following these structural characteristics in the extracellular region, a typical transmembrane domain is seen in the 23 amino acids from the 233rd Gly to the 255th Leu. In the intracellular region that follows, the 271st and 273rd Pro residues are Box-1 consensus sequence (PXP motif) conserved in other hemopoietin receptor members, and these are thought to be deeply involved in signal transduction. Thus, mNR8y adequately satisfies the characteristics of hemopoietin receptor members.

[0187] On the other hand, for mNR8 β , among the structural characteristics for the above-mentioned extracellular region, the exon sequence encoding the Pro-rich region has been skipped by selective splicing, and directly joins the next exon encoding the WS motif. However, the WSXWS-Box sequence has been excluded from the reading frame by frame shift, and after coding up to 144thLeu, the translation frame completed the next stop codon. Thus, a soluble hemopoietin receptor-like protein that does not have a transmembrane domain is encoded.

Example 8: Expression analysis of mouse NR8 gene

8-1) Analysis of mouse NR8 gene expression by the RT-PCR method

[0188] To analyze the distribution and mode of NR8 gene expression in each mouse organ, the mRNA was detected by RT-PCR analysis. As primers for this RT-PCR analysis, NR8-SN1 primer (SEQ ID NO: 9) was used as the sense side (downstream direction) primer, and NR8-AS1 primer was used as the antisense side (upstream direction) primer. Mouse Multiple Tissue cDNA Panel (Clontech #K1423-1) was used as the template. Advantage cDNA Polymerase Mix (Clontech #8417-1) and the Perkin Elmer Gene Amp PCR System 2400 Thermalcycler were used for PCR. The target genes were amplified by the PCR reaction under the cycle condition given below.

[0189] PCR conditions were 94°C for 4 min, 5 cycles of "94°C for 20 sec, 72°C for 1 min," 5 cycles of "94°C for 20 sec, 70°C for 1 min," 24 cycles of "94°C for 20 sec, 68°C for 1 min," 72°C for 3 min, and completed at 4°C.

[0190] The results of RT-PCR are shown in Fig. 18. The NR8 gene was strongly detected in the testis and day 17 embryo, and a constitutive gene expression was seen in all mouse organs and in all mouse tissue-derived mRNA analyzed. By detecting the expression of the house keeping gene G3PDH under the above-mentioned PCR conditions using the mouse G3PDH primer for all the templates used in the analysis, it has been verified beforehand that the number of copies of template mRNA has been normalized (standardized) between samples. The detected RT-PCR product size herein was 320 bp, and this coincides with the size calculated by the determined nucleotide sequence. Therefore, it was thought to be the product of the mouse NR8 specific PCR amplification reaction. To further verify this, the PCR product amplified in the day 17 embryo was subcloned to pGEM-T Easy vector according to 7-2), and the nucleotide sequence was analyzed. The result verified that the PCR product could be a partial nucleotide sequence of mouse NR8, and the possibility that it might be the product of a non-specific PCR amplification was denied.

8-2) Analysis of mouse NR8 gene expression by Northern blotting

[0191] In order to analyze NR8 gene expression in each mouse organ, and with the objective of identifying the NR8 transcription size, gene expression analysis by the Northern blotting method was conducted. Mouse Multiple Tissue Northern Blot (Clontech #7762-1) was used as the blot. Among the 5'-RACE products obtained in 7-4), the mNR8β cDNA fragment was used as the probe. The probe was radiolabeled with [α-32P] dCTP (Amersham, cat#AA0005) using Mega Prime Kit (Amersham, cat#RPN1607). Express Hyb Hybridization Solution (Clontech #8015-2) was used for hybridization. After a prehybridization at 68°C for 30 min, the heat-denatured labeled probe was added, and hybridization was conducted at 68°C for 16 hr. After washing under the following conditions, the blot was exposed to Imaging Plate (FUJI #BAS-III), and a mouse NR8 specific signal was detected by the Image Analyzer (FUJIX, BAS-2000 II).

[0192] Washing conditions were: (1) 1x SSC/0.1% SDS, at room temperature for 5 min; (2) 1x SSC/0.1% SDS, at 50°C 30 min; and (3) 0.5x SSC/0.1% SDS, at 50°C 30 min.

[0193] As a result, as shown in Fig. 19, a strong expression was seen in the mouse testis only, and no gene expression of the same gene was detected in other organs. Here, there is a difference between the results of RT-PCR analysis and Northern blot analysis. Since the detection sensitivity of the Northern method is much lower than RT-PCR, it is thought that mRNA with low expression levels could not be detected. However, results of both analyses coincide in the point that a strong gene expression was detected in the testis. Also, the size of the detected transcript was about 4.2 kb. [0194] Although there was a deviation of the expression levels in each mouse organ analyzed by the Northern method and RT-PCR, the gene expression was widely distributed, being detectable in all the organs analyzed especially

when using RT-PCR. This result contrasts with the human NR8 gene in which the expression was strong only in immunocompetent tissues, hemopoietic tissues, and specific leukemic cell lines, and the significance of this expression is extremely interesting. This means namely the possibilities that in mouse, the NR8 molecule not only is involved in systemic hemopoietic functions, or in immunological responses, and hemopoiesis, but also may be involved in various physiological regulatory mechanisms of the body. Namely, its ligand may be able to function as a hormone-like factor.

Example 9: Isolation of the NR8 mouse genomic gene by plaque screening

The present inventors analyzed the genomic structure of mouse NR8 gene and performed plaque hybridization against the mouse genomic DNA library. 129SVJ strain Genomic DNA (Stratagene #946313) constructed in Lambda FIX II was used as the library. This genomic library of approximately 5.0 x 10^5 plagues was developed and blotted to a Hybond N(+)(Amersham #RPN303B) charged nylon membrane to perform primary screening. NR8 β cDNA fragment of 5'-RACE products obtained in 7-4) was used as the probe. The probe was radiolabeled with $\{\alpha^{-32}P\}$ dCTP prepared as above-mentioned in 8-2) using the Mega Prime Kit. Express Hyb Hybridization Solution was used for hybridization, and after a prehybridization at 65°C for 30 min, a heat-denatured labeled probe was added, and hybridization was done at 65°C for 16 hr. After washing under the following conditions, the membrane was exposed to an X-ray film (Kodak, cat#165-1512) to detect mouse NR8 positive plaques.

[0196] Washing conditions were: (1) 1x SSC/0.1% SDS, at room temperature for 5 min; (2) 1x SSC/0.1% SDS, at 58°C 30 min; and (3) 0.5x SSC/0.1% SDS, at 58°C 30 min.

[0197] As a result, positive, or pseudo-positive 16 independent clones were obtained. When a secondary screening was similarly conducted against these 16 clones obtained by the primary screening, the inventors succeeded in isolating NR8 positive, nine independent plaque clones.

Industrial Applicability

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[0198] The present invention provides a novel hemopoietin receptor protein "NR8," and the encoding DNA. The present invention also provides, a vector into which the DNA has been inserted, a transformant harboring the DNA, and a method of producing a recombinant protein using the transformant. It also provides a method of screening a compound or a natural ligand that binds to the protein. The NR8 protein of the invention is thought to be related to hemopoiesis, and therefore, is useful in analyzing hemopoietic functions. The protein would also be applied in the diagnosis and treatment of hemopoiesis-associated diseases.

[0199] Since the expression of mouse NR8 gene was widely distributed in mouse organs, mouse NR8 protein would be involved in various physiological regulatory mechanisms of the body, including the above-mentioned hemopoiesis. Furthermore, by using mouse NR8 protein, it is possible to isolate first the mouse NR8 ligand, and next, the human homologue of the NR8 ligand using the conserved structure of the mouse NR8 ligand. Specifically, after determining the nucleotide sequence of mouse NR8 ligand cDNA, an oligonucleotide primer is designed on this sequence, and using this to conduct cross PCR using the human-derived cDNA library as the template, human NR8 ligand cDNA can be obtained. Alternatively, human NR8 ligand cDNA can be obtained by conducting cross hybridization against human-derived cDNA library using mouse NR8 ligand cDNA as the probe. It is also possible to analyze biological function of the NR8 receptor protein by creating a mouse NR8 gene-deficient mouse using the mouse NR8 gene.

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	roh	Ile	гце	95	AST	YZII	116	ım		GIN	5er	GIY	Asn		Ser.	Gin	
30				90					100					105			
	GAG	TGT	സാ	AGC	TTT	CTC	ርጥር	ርርጥ	CAC	ACC	AAC	T 00	CAC	0.40	444	0.07	000
	GIn	Cys	Glv	Ser	Pho	T.en	Len	Ala	Clu	700 702	AAU Two	202	Clu	CAU	AAA T	GC I	809
	014		110	UCI .	ILC	DÇU		115	VIU	DC1.	r)3			ata	LYS	AIA	
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	Asp	Leu	Ser	Glv	Leu	l.vs	ī.vg (Cve	Len i	Pro i	Don	Dno	Dog	684 61 4	guu Vai	Dno	857
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20	ata	0+0	- ant	ero er		a t a			+		هـــــــــــــــــــــــــــــــــــــ						004
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5		130	0				13	5				140)				
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	CC.	t gc	C CCC	CLEC	S ate	CLE	, 88 . T	ggg	: aag	cti	t cau	g tai	ga	g cti	cau	g tac	480
			1 PD	e lyl	r net			3 (11)	Lys	3 Let			· 611	ı Lei	1 611	1 Tyr	
10	149) _.				150	,				158	•				160	
	ags	2 880	: CEE	7 PP	L gac	ccc	t.gr	r rct	et.	r apt	cce	7 900	r sagre	999	, cto	g atc	528
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25		•	195	ı				200			•		205	1			
	tee	tec	റാത		900	+ 00	2 ort	ora o	+ 00	ant	500	000	~+ ~	n+-	111	cag	670
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	000	000	220	000	004	an a		ata	.+-	-+-				4			010
								ctc Leu									816
	110	110	ща	260	பர்க	NOP	ıщ	neu	265	116	Set.	WLR	IIII.	270	GIU	AST	
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· -														•		•	
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	Phe	Tyr	Pro :	Ser A	Asp	lle	Ala	Val (3lu 1	ſrp (Hú S	Ser A	lsn (Gly (Gln i	Pro '

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15	· ·	171L	01-	*	: 	*		. T	. C				•.				
	lyr			гÀЗ	s ser	reu	455			rrc	9 613	460			,		.:.
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ю	Gly '																30
				20	-				25	·				30			
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5	Val																
			• ;														
ю	ctt a																192
•	Leu 1		[rp	Gln	Asp	Gln		Glu	Glu	Leu	Lys		Ğlu	Ala	Thr	Ser	
		50					55					60					

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25	aag :	aag Lys 130:	Cys	ctc Leu	ect Pro	cct Pro	ccc Pro 135	cct Pro	gga Gly	gtt Val	ccg Pro	caa Gln 140	aga Arg	ctc Leu	gag	cta Leu	432
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40	Gly	Trp	Gly	Cys 20	Pro	Asp	Leu	Val	Cys 25	Tyr	Thr	Asp	Tyr	Leu 30	Gln	Thr
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40	Gln					-00		-0 0	-0-0	60~~	0 00	0000	OB C C	400			310
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Claims

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- 1. A protein comprising the amino acid sequence from the 1st amino acid Met to the 361st amino acid Ser of SEQ ID NO: 1, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 361st amino acid Ser of SEQ ID NO: 1.
- 2. A protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 3, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 3.
- 15 3. A protein comprising the amino acid sequence from the 1st amino acid Met to the 237th amino acid Ser of SEQ ID NO: 5, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 237th amino acid Ser of SEQ ID NO: 5.
 - 4. A protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 7, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added and/or substituted with another amino acid, and being functionally equivalent to the orotein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 7.
 - 5. A protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 19, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 19.
 - 6. A protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 21, or a protein comprising a modified amino acid sequence of said amino acid sequence in which one or more amino acids have been deleted, added and/or substituted with another amino acid, and being functionally equivalent to the protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 21.
- 7. A protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 2, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 361st amino acid Ser of SEQ ID NO: 1.
 - 8. A protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 4, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 3.
 - A protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 6, which is functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 237th amino acid Ser of SEQ ID NO: 5.
 - 10. A protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 8, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 7.
- 55 11. A protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 20, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 144th amino acid Leu of SEQ ID NO: 19.

- 12. A protein encoded by a DNA hybridizing to a DNA comprising the nucleotide sequence of SEQ ID NO: 22, said protein being functionally equivalent to a protein comprising the amino acid sequence from the 1st amino acid Met to the 538th amino acid Ser of SEQ ID NO: 21.
- 13. A fusion protein comprising the protein of any one of claims 1 to 12 and another peptide or polypeptide.
 - 14. A DNA encoding the protein of any one of claims 1 to 13.
 - 15. A vector comprising the DNA of claim 14.

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- 16. A transformant harboring the DNA of claim 14 in an expressible manner.
- 17. A method of producing the protein of any one of claims 1 to 13, comprising the step of culturing the transformant of claim 16.
- 18. A method of screening a compound that binds to the protein of any one of claims 1 to 13 comprising the steps of:
 - (a) contacting a test sample with the protein of any one of claims 1 to 13; and
 - (b) selecting a compound that comprises an activity to bind to the protein of any one of claims 1 to 13.
- 19. An antibody that specifically binds to the protein of any one of claims 1 to 12.
- 20. A method of defecting or measuring the protein of any one of claims 1 to 13 comprising the steps of contacting a test sample presumed to contain said protein with the antibody of claim 19, and detecting or measuring the formation of the immune complex between the antibody and the protein.
- 21. A DNA specifically hybridizing to a DNA comprising a nucleotide sequence of any one of SEQ ID NOs: 2, 4, 6, 8, 20, and 22 to 27 comprising at least 15 nucleotides, and comprising at least 15 nucleotides.

33	481	331	232	243	238	438	242	438	329	255	247	244	229	319
41032	4	က	2	7	2	4	2	4	n	2	7	7	7	
SLLPLEF RKDSSYELQVRAGPMPGSSYQGTWSEWSDPV1FQTQSEGRCEAGMDTPLL	LELRPRSRYRLQLRAR-LNGPTYQGPWSSWSDPTRVETATE	SLLVDSILPGSSYEVQVRGKRLDGPGIWSDWSTPRVFTTQ	DTQYEFQVRVKPLOGEFT-TWSPWSQPLAFRIK	TLLORKLOPAAMYEIKVRSIPDHYFKGFWSEWSPSYYFRIPEINNSSGEMDPILL	T <u>LGPEHLMPSSTY</u> VAR <u>VR</u> TRLA <u>PGS</u> RLSGRPSK <u>WS</u> PE <u>V</u> CWDSG	TGYNGIWSEWSEARSWDIES	N <u>L</u> EPKL <u>F</u> LPN <u>SIYAARVR</u> TRLSA <u>GSS</u> LSGRPSR <u>WS</u> PE <u>V</u> HWDSQ	QLEPDISYCARVRVKPI-SDYDGIWSEWSNEYTWTI	SKYDVQVRAAVSSMCREAGLWSEWSQPI	YTGOWSEWSOPVCFQ	RGRTRYTFAVRAR-MAEPSFGGFWSAWSEPVSLLIPSD	SLPSVDGOKRYTFRVRSRFNPLCGSAQH-WSEWSHPI	LCPLEMNVAGEFOLRREGLGSGGSSWSKWSSPV	LDLKPFTEYEFGISSKLHLYKGSWSDWSESLRAQIPEE
40004	442	292	201	189	196	419	200	404	302	241	211	209	197	282
2	HTPOR	hobr	hIL2Rb	hIL7R	hGM-CSFRb		mIL3Rb		hIL5Ra	h119R	hEPOR	hIL2Rr	hIL12R	hIL12Rb

[Query: 39181-39360]

NR8 ·	39233	HQVKPAPPFN-VTVTFSGQYNISWRS-DYEDP-AFYMLKGKLQY 39355
hIL6Ra	214	LOPDPPANITVTAVAR-NPRWLSVTWODPHSWNSSFYRLRFELRY 257
hgp130	218	YK <u>VKPNPPHN</u> L-S <u>V</u> INSEELSSILKLTWT-NPSIKSVII <u>LK</u> YNIQY 261
rOBRb	234	VKPDPPLGLRHEVTDDGNLKISWDS-QTKAP 263

[Query: 42301-42480]

NR8	42307	VPSPERFFMPLYKGCSGDFK 42366	
mIL9R	305	IPSPEAFFHPLYSVYHGDFQ 324	
hIL9R	305	VPSPAMFFOPLYSVHINGNFQ 324	

Figure 3

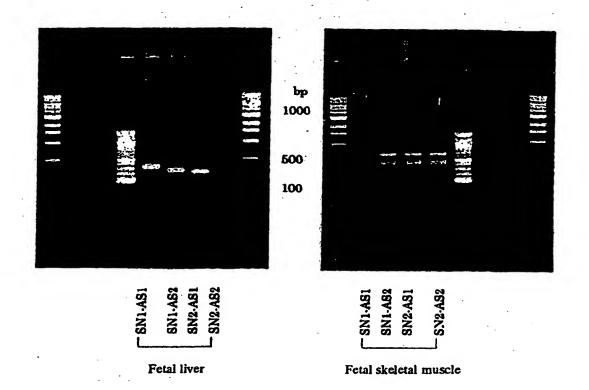
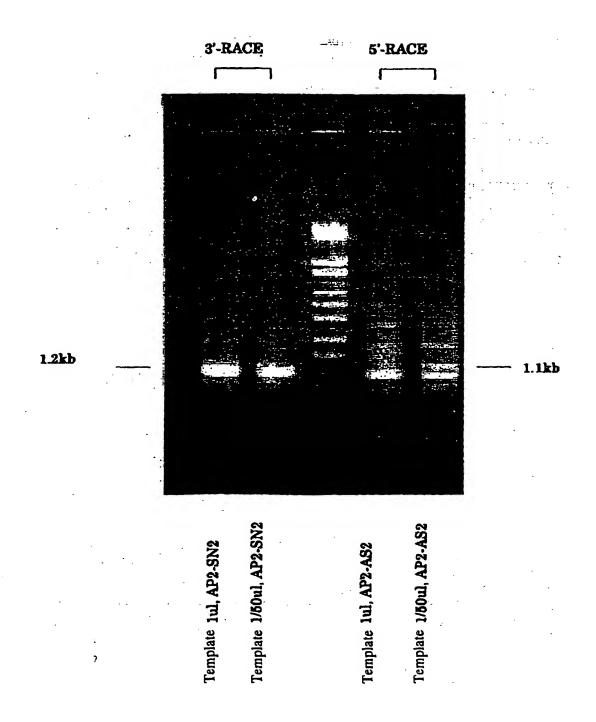


Figure 4



10	20	30	40	50	60	70	80
GGCAGCCAGCGGCC	TCAGACAGACCC	ACTGGCGTC	TCTCTGCTG	AGTGACCGTA	AGCTCGGCGT	CTGGCCCTCT	GCCTGC
90	100	110	120	130	140	150	. 160
CTCTCCCTGAGTGT	TGGCTGACAGCCA	CGCAGCTET	GICTGICTE	TCTGCGGCCC	STECATCCCT	GCTGCGGCCG	CCTGGT
•							
170	180	190	200	210	220	230	240
ACCITCCTTGCCGT	CICILICCICIE	TCTGCTGCT	CTGTGGGAC	ACCTECCTEG	AGGCCCAGCT	GCCCGTCATC	AGAGTG
250	260	270	280	290	300	310	320
ACAGGTCTTATGAC	AGCCTGAT IGGT	SACTUGUGU	IGGG1G1GG	ATTOTCACCO	CAGGCCTCTG	CCIGCITICIO	CAGACC
330	240	350	360		200	200	:400
CTCATCTGTCACCC	340 CCACCCTCAACC			370	380	390	400
CICATCIBICACC	TCMCOC 1 GAVIO	WIGO I GOOM	·	BULLIA I LAUA		UNUNCUENN I	JUNI I
410	420	430	440	450	460	470	480
CTGAGAAAGAAGCC							
					AAP		LL
490	500	510	520	530	540	550	560
TCCAGGGAGGCTGG						CATCCTGGAA	ATGTGG
QGGW	GCPDI			YLQT			H W
570	580	590	600	610	620	630	640
AACCTCCACCCCAG							
	TLTL				K D E A		SL
650 CCACAGGTCGGCCC	660	670	680 ACACCTCCC	690	700	710	720
H R S A H							I F
730	740	750	760	770	780	790	800
TCAGTGTCAACATC	* * * *						
	T D Q S			C G S F			K P
810	820	830	840	<i>8</i> 50	860	870	880
GCTCCCCCTTTCAA	CGTGACTGTGACC	TTCTCAGG	ACAGTATAA	TATCTCCTGG	CGCTCAGATT	ACGAAGACCC	IGCCTT
APPFN	VTVT	FSG	QYN	ISW	RSDY	E D P	A F
890	900	910	920	930	940	950	960
CTACATGCTGAAGG	GCAAGCTTCAGT/						EAGAA
YHLKG				R G D		· ·	R K
970	960	990	1000	1010 -	1020	1030	1040
AGCTGATCTCAGTG							
	DSRSV			E F R K) V
1050		070	1080	1090	1100	1110	1120
CGGCAGGCCCAT	_						
RAGPH	PGSS	TUU	TWS	EWSI	PVI	FQT	QS

AAAAAAAAAAAA

1180 1190 AGAGGAGTTAAAGGAAGGCTGGAACCCTCACCTGCTTCTCCTCCTCCTGCTTGTCATAGTCTTCATTCCTGCCTTCTGGA EELKEGWNPHILLLLLVIVFIPAFWS **ECCTGAAGACCCATCCATTETGGAGGCTATGGAAGAAGATATGGGCCGTCCCCAGCCCTGAGCGGTTCTTCATGCCCCTG** LKTHPLWRLWKKIWAVPSPERFFHPL TACAAGGECTECAGCGGAGACTTCAAGAAATGGGTGGGTGCACCCTTCACTGGCTCCAGCCTGGAGCTGGGACCCTGGAG YKGCSGDFKKWVGAPFTGSSLELGPWS PEVPSTLEVYSCHPPSSPVECDFTSPG DEGPPRSYLROWVVIPPPLSSPGPQA-AGCTAATGAGGCTGACTGGATGTCCAGAGCTGGCCCAGGCCACTGGGCCCTGAGCCAGAGACAAGGTCACCTGGGCTGTGA S * * TECCTETESCCTTESCATAATGCCCATGGTACTCCATGCATTCACCTCCCTGTGCATGTCTGGACTCACGGAGCTCACC

10	20	30	40	50	60	70	80
GECAGCCAGCG	GCCTCAGACAGAC	CCACTGGCGT	CTCTCTGCT	GAGTGACCGTA	AGCTCGGCGT	ICTESCCCTCT	
			•				
. 90	100	110	120	130	140	150	160
CTCTCCCTGAG	TETEECTEACAGE	CACGCAGCTE	TGTCTGTCTC	STCTGCGGCCC	ETECATOCCI	FECTECESCO	CCTGGT
		re Fig. 1					
170	180	-190	200	210	220	230	240
ACCTTCCTTGC	CETCTCTTTCCTC	TETCTECTEC	TCTGTGGGA	CACCTGCCTGG	AGGCCCAGCT	FECCEST CATE	
•	* **	` : 	· · .				
250	260	270	280	290	300	310	320
ACAGGTCTTAT	GACAGCCTGATTG	GT6ACTCGGG	CTGGGTGTGG	SATTCTCACCC	CASSCCTCTC	SCCTGCTTTCT	CAGACC
	• :	****	•				
330	340	350	360	370	380	390	400
CTCATCTGTCA	CCCCACGCTGAA	CCCAGCTGCC	ACCCCCAGA	VECCCATCAGA	CTGCCCCCAG	CACACGGAAT	GGATTT
410	420	430		· 450		470	480
CTEAGAAAGAA	GCCGAAACAGAAG	GCCCGTGGGA					
			M P	RGW	AAP	LLLL	LL
		·					
490	500	510	520	530	540	550	560
	IGGGGCTGCCCCG						
0 6 G I	e G C P D	LVC	YTD	YLQT	A I C	ILE	M A
570	500	500					
570		590				630	640
	CAGCACGCTCACC S T L T						
нене	5 1 L !		ז גיי ט	FFL	KUEA	1 1 2 6	2 L
650	660	670	680	690	700	710	720
500	CCACAATGCCAC					•	
	HNAT						
n k y. n						WPT	
730	740	750			780	790	800
	TCACABACCAGT						
	TDQS						
	SOTSI						
810	820			850		870	880
	CTCAGTGGACTCA					• • •	
	LSGL						,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,
	SVDS						1 0
890				930		950	960
•••	GGCCCATGCCTGG				-		
?							

Figure 8

1000 970 980 990 1010 1020 1030 1040 CAGTCAGAGGAGTTAAAGGAAGGCTGGAACCCTCACCTGCTGCTTCTCCTCCTGCTTGTCATAGTCTTCATTCCTGCCTT Q S E E L K E G W N P H L L L L L L V I V F I P A F 1050 1060 1070 1060 1090 1100 CTGGAGCCTGAAGACCCATCCATTGTGGAGGCTATGGAAGAAGATATGGGCCGTCCCCAGCCCTGAGCGGTTCTTCATGC W S L K T H P L W R L W K K I W A V P S P E R F F N P 1140 1150 1160 1170 1180 LYKGCSGDFKKWVGAPFTGSSLELGP 1210 1220 1230 1240 1250 1260 TGGAGCCCAGAGGTGCCCTCCACCTGGAGGTGTACAGCTGCCACCCAGCAGCCCTGTGGAGTGTGACTTCACCAG W S P E V P S T L E V Y S C H P P S S P V E C D F T S 1290 1300 1310 1320 1330 1340 1350 1360 P G D E G P P R S Y L R Q W V V I P P P L S S P G P Q 1370 1380 1390 1400 1410 1420 1430 1440 AGGCCAGCTAATGAGGCTGACTGGATGTCCAGAGCTGGCCAGGCCACTGGGCCCTGAGCCAGAGACAAGGTCACCTGGGC A S * * 1450 1460 1470 1480 1490 1500 1510 1520 1540 1550 1560 ...1570 1530 1580 1590 1600 1620 1630 1640 1610 1650 1660 1670 GCACGTGCCTGTGGGCCTGGGATAATGCCCATGGTACTCCATGCATTCACCTGCCCTGTGCATGTCTGGACTCACGGAGC

1770 1780 AAAAAAAAAAAAAAA

1690

1700 1710

1720

1730

1740

10	20	30 ·	40	50	60	70	80
GGCAGCCAGCGGCC	TCAGACAGA	CCACTGGCGT	CTCTCTGCT	PATTGACCGTA	AGCTCGGCG1	CTGGCCCTCT	GCCTGC
90	100	110	120	130	140	150	160
CTCTCCCTGAGTGT	GGCTGACAGO	CACGCAGCTO	TGTCTGTCTG	TCTGCGGCCC	GTGCATCCCT	GCTGCGGCCG	CCTGGT
					-		
170	180	190	200	210	220	230	240
ACCITICATION	CTCTTTCCTC	TGTCTGCTGC	TCTGTGGGA	CACCTGCCTGG	AGGCCCAGCT	GCCCGTCATC	
250	260	270	280	290	300	310	320
ACAGGTCTTATGAC	AGCCTGATTO	EGTGACTCGGE	CTGGGTGTG	ATTETCACCE	CAGGCCTCTG	CCTCCTTTCT	
						,	
330	340 . 🚽	350	360	. 370	- 380	390	400
CTCATCTGTCACCC		<i>.</i> .				CACACGGAAT	
•					-		
410	420	430	440	450	460	470	480
CTGAGAAAGAAGCC	GAAACAGAAG	GCCCGTGGGA	GTCAGCATGC	CCCCTCCCTC	GGCCGCCCCC	TTGCTCCTGC	TECTEC
			M P	RGW	AAP	LLLL	LL
490	500	510	520	530	540	550	560
TCCAGGGAGGCTGG	GECTECCCC	ACCTCGTCTG	CTACACCGAT	TACCTCCAGA	CEGTCATCTE	CATCCTGGAA	ATGTGG
9 6 6 W	GCPD	LVC	YTD	YLQT	VIC	ILE	M W
570	580	590	600	610	620	630	640
AACCTCCACCCCAG							CAGCCT
NLHPS	TLT	LTWO	DQY	EEL	KDEA	TSC	SL
650	660	670	680	690	700	710	720
CCACAGGTCGGCCC							
HRSAH							I F
730	740	750	760	770	780	790	800
TCAGTGTCAACATC							
SVNI	_		_				
810	820	830	840	850	860	870	880
GCTCCCCCTTTCAA							
APPFN							A F
890	900	910	920	930	940	950	960
CTACATECTGAAGGC							
YMLKG						•	
970	980	990	1000	1010 -	1020	1030	1040
AGCTGATCTCAGTGC							
			L P L) V
1050	1060	1070	1080	1090	1100	1110	1120
COGCAGGCCCATC							
RAGPH	7 6 5 9	. T D G	1 # 5	2 M 4) P V T	F 0 T	U 5

1130	1140				1180		
AGAGGAGTTAAAG							TTCTGGA
EELKE	e a n	PHL	LLLi	LLV	IVF	IPAF	W S
1210	1220	1230	1240	1250	1260	1270	1280
GCCTGAAGACCCAT	CCATTGTGG	AGGCTATGGA	AGAAGATATG	GGCCGTCCCC	AGCCCTGAGC	GGITCTTCAT	GCCCCTG
LKTH							
1290	1300	1310	1320	1330	1340	1350	1360
TACAAGGGCTGCAG	CGGAGACTT	CAAGAAATGG	GTGGGTGCAC	CCTTCACTGG	CTCCAGCCTG		DARTTY
YXGCS							
1370	1380		1400		1420	1430	1440
CCCAGAGGTGCCCT	CCACCCTGG						
PEVPS	TLE	VYS	CHP	PRSP	AKR	1 0 1	TFI
1450	1460	1470	1480	1490	1500	1510	1520
TACAAGAACCAGCA	GAGCTEGTGE	AGTCTGACG	TGTGCCCAA				
QEPA							
					<u> </u>	<u> </u>	<u> </u>
1530	1540	1550	1560	1570	1580	1590	1600
TCAGCTTACAGTGA	GGAGAGGGAT	CGGCCATACE					
SAYSE	ERD	RPY	LVS	I D T	VTV	LDAE	
					······································		
1610	1620	1630	1640	1650	1660	1670	1680
ATECACCTEGCCCT	GCAGCTGTGA	GGATGACGGC	TACCCAGCC	CTGGACCTGG			
CTWPC							
1690	1700	1710	1720	1730	1740	1750	1760
TAGAGGACCCACTC	TTGGATGCAG	GGACCACAGT	CCTGTCCTGT	IGGCTGTGTCT	CAGCTGGCA	GCCCTGGGCT	AGGAGGG
EDPL							
1770	1780	1790	1800	1810	1820	1830	1840
CCCCTGGGAAGCCT	CCTGGACAGA	CTANAGCCAC	CCCTTGCAGA	TGGGGAGGAG	TEEGCTGGG	GACTGCCCT	GGGGTGG
PLGSL	LDR	LKPP	LAD	GED	WAGG	LPW	GG
						•	
1850	1860	1870	1880	1890	1900	1910	1920
CCGGTCACCTGGAG	EGGTCTCAGA	GAGTGAGGCG	GECTCACCCC	TEECCECCT	GGATATGGAC	CACGTTTGAC	
0 0 0 0				_			
K O F U U	V S E	SEA	GSPL	AGL	D M D	TFD	SGF
KSFUU	Y S E	SEA	GSPL	AGL	D M D	T F D	S G F
1930	V S E	S E A (<u>G S P L</u> 1960	. A G L 1970	D M D	T F D :	2000
	1940	1950	1960	1970	1980	1990	2000
1930	1940 FGCAGCAGCC	1950 CTGTGGAGTG	1960 TGACTTCACC	1970 AGCCCCGGGG	1980 ACGAAGGACO	1990 CCCCCGGAG	2000 CTACCTC
1930 TTGTGGGCTCTGACT	1940 FGCAGCAGCC	1950 CTGTGGAGTG	1960 TGACTTCACC	1970 AGCCCCGGGG	1980 ACGAAGGACO	1990 CCCCCGGAG	2000 CTACCTC
1930 TTGTGGGCTCTGACT	1940 FGCAGCAGCC	1950 CTGTGGAGTG	1960 TGACTTCACC	1970 AGCCCCGGGG	1980 ACGAAGGACO	1990 CCCCCGGAG	2000 CTACCTC
1930 TTGTGGGCTCTGACT V G S D C	1940 GCAGCAGCCC S S P 2020	1950 CTGTGGAGTG V E C	1960 TGACTTCACC D F T 2040	1970 AGCCCCGGGG S P G D 2050	1980 ACGAAGGACO E G P 2060	1990 CCCCCGGAG P R S 2070	2000 CTACCTC Y L 2080
1930 TTGTGGGCTCTGACT	1940 IGCAGCAGCCC S S P 2020 CATTCCTCCGC	1950 CTGTGGAGTG V E C 2030 CCACTTTCGAG	1960 TGACTTCACC D F T 2040 SCOCTGGACC	1970 AGCCCCGGGG S P G D 2050 CCAGGCCAGC	1980 ACGAAGGACO E G P 2060 TAATGAGGCT	1990 CCCCCGGAG P R S 2070	2000 CTACCTC Y L 2080

	2090	2100	2110	2120	2130	2140	2150	2160
CTGG	CCAGGCCAC	TEGECCCTE	GCCAGAGACA	AGGTCACCTE	GECTETEATE	TGAAGACACC	TGCA6CCTTT	EGTCTCC
	2170	2180	2190	2200	2210	2220	2230	2240
TGGA	TGGGCCTTT	EAGCCTGATE	STITACAGTGI	CIGIEIGIGI	GTGCATATGT	<u> दाहादादाद</u> ्	ATATGCATGT	etetete
	2250	2260	2270	2280	2290	2300	2310	2320
TGTG	TGTGTCTTA	GGTGCGCAGT	IEGCATGTCCA	CETETETETE	ATTECACETE	CCTGTGGGCC	TGGGATAATG	CCCATGG
	2330	2340	2350	2360	2370	2380	2390	2400
TACT	CCATGCATT	CACCTGCCCT	GTGCATGTCT	EGACT CACGG	AGCTICACCCA	TGTGCACAAG	TETECACAGT	AMACETE
	2410	2420	2430	2440	2450	2460	2470	2480
THE	TGGTCAACA						AA .	

Figure 12

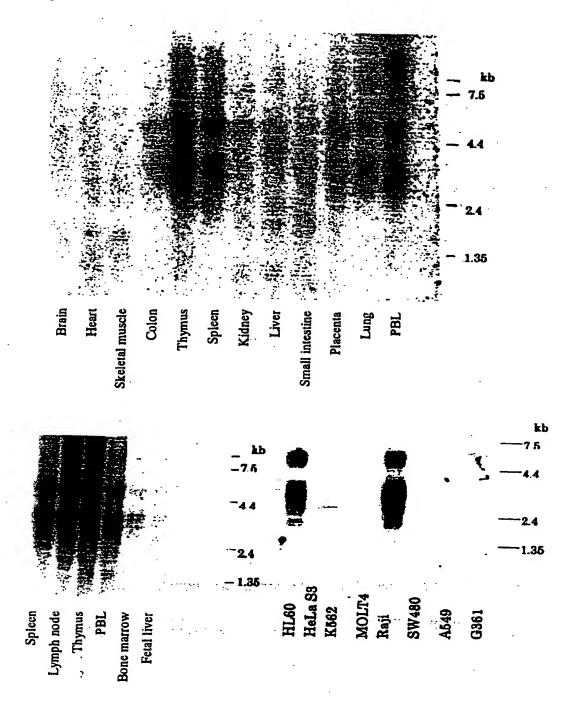
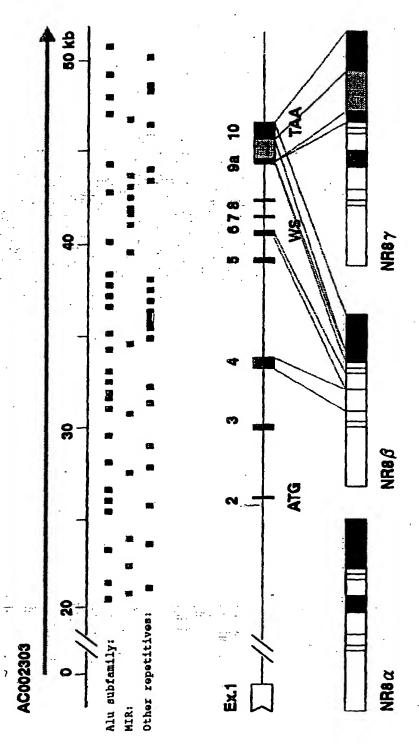
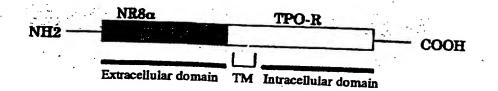


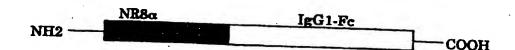
Figure 13



pEF-NR8/TPO-R



pEF-NR8/ IgG-Fc



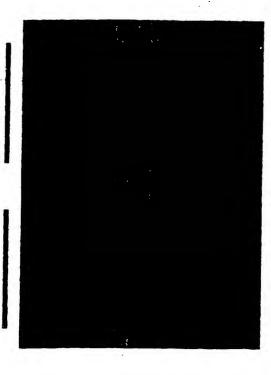
pEF-BOS/NR8b FLAG



Pigure-15

Mouse Brain cDNA

Mouse Testis cDNA



100 bp Ladder
[NR8-SN2 / NR8-AS2]

[NF18-SN2 / NR8-AS1]

[NR8-SN1 / NR8-AS2]

[NR8-SN1 / NR8-AS1]

100 bp Ladder

100 bp Ladder

[NR8-SN2 / NR8-AS2]

[NR8-SN2 / NR8-AS1]

[NR8-SN1 / NR8-AS2]

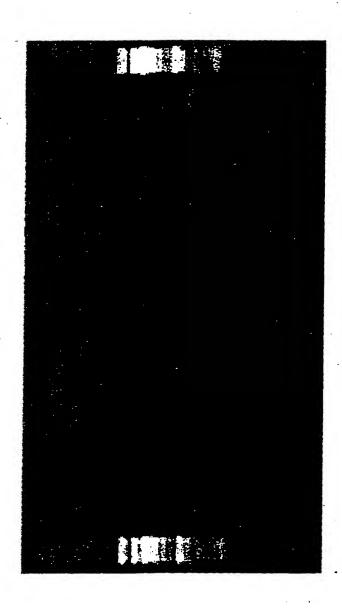
[NR8-SN1 / NR8-AS1]

100 bp Ladder

hnr8beta	upper apul	LILL COSSIGN	PRIATERINA	GLAXCO SECTION	40
mmr8beta	HUNGRALISED		PROVENCEDYS	2717767372940	40
hnrebeta		ñao o 111223		AUDICALIZAÇÃ	80
mnrsbeta	Manusa Pre 20	ACE CONTRACT	EASSES ONES	ABULTUATED	80
hnrsbeta	CHONTERNA	BINTE CONTRACTOR	esessore	SENLARGESE	120
mnrsheta	SHARREN	0.0 september 10	oscier poet	SEPRESESE	120
hnrsbeta	PPZHUSGURE	chimpresure	PLYE		144
mersbeta	ECHOPSELES.	CEPERRGYES	RDEEL.		144

hnrsg mnrsg			Didayanaya Didayanaya	***************************************	40 40
hnreg mnreg			EATRICSUME PATRICSUME		80
hersg mersg		- And the contract of the contract of the con-	CSENIZSONES ESCNIPONCE		120 120
hersg mersg	ATA: A. MANONE MANAGEMENT AND		DYEDPATYME	Westerlebensen Same	160 160
hNRSG mNRSG	RINGEDEMANS	PHRWLISVOS	REVELLELER REVELLER	BEDSEVELON	200 200
hnrsg mnrsg			end (desployable Particologic State		240 240
hersc mersc		,	Bulgaret kan Referense	A4444-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-	280 280
hersg rersg	·///	A 1 1 2 1 2 1 2 1 2 1 2 1 2 1 2 1 2 1 2	6275251252 62763535451	Constitution and the second	320 320
hNRSG mNRSG	PERSONAL SECTION	***********************	VESDGVETUS VESDGVETUS	prostational protection of	360 360
hnreg mnreg	***************************************	MODUSLIJE JE KODNSTILIJE	SADABGREDN SADBSCRASS	PCSEEDDGYE PCSEEDDGYE	400 400
LMRSG mnrsg	- WACOO GOLDEN TOWN CO. WALLES	***************************************	ACONCRESCE ACONCRESCES	The same of the sa	440 440
hereg mereg	CANADA TARANTA MANAGAMAN	The second secon	CGMACGRAP	***************************************	480 480
hnrsg marsg	Care of the Second of same the same of		SHVEGDETSE SHVEGDETSE	Charles and an arrangement	520 520
heirsg mersg	ROMVVIPPEL ROMVVIPPEL	***************************************			538 538

Figure 18



100 bp Ladder

E17-day

E15-day

E11-day

E7-day

Testis

Kidney

Skeletal muscle

Liver

Lung

Spleen

Brain

Heart

100 bp Ladder

Figure 19



Testis

Kidney

Skeletal muscle

Liver

Lung

Spleen

Brain

Heart



Mouse Beta-actin (2.0kb and 1.8kb)

INTERNATIONAL SEARCH REPORT International application No. PCT/JP99/03351 CLASSIFICATION OF SUBJECT MATTER C07K14/715, C07K19/00, C12N15/12, C12N5/10, C12P21/02, G01N33/50, C07K16/16/28, G01N33/53, C12Q1/68 According to International Patent Classification (IPC) or to both national classification and IPC **B. FIELDS SEARCHED** Minimum documentation searched (classification system followed by classification symbols) Int.Cl⁵ C07K14/715, C07K19/00, C12N15/12, C12N5/10, C12P21/02, G01N33/50, C07K16/16/28, G01N33/53, C12Q1/68 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) GenBank/EMBL/DDBJ/GeneSeq, SwissProt/PIR/GeneSeq C. DOCUMENTS CONSIDERED TO BE RELEVANT Category* Citation of document, with indication, where appropriate, of the relevant passages Relevant to claim No. J. Biol. Chem. 271[23] (1996) Robb L. et al., "Structural Analysis of the Gene Encoding the Murine 1-21 Interleukin-11 Receptor α-Chain and a Related Locus* p.13754-13761 A 1-21 Proc. Natl. Acad. Sci. USA 93 (1996) Gainsford T. et al., "Leptin can induce proliferation, differentiation, and functional activation of hemapoietic cells p.14564-14568 A 1-21 Proc. Natl. Acad. Sci. USA 93 (1996) Hilton D.J. et al., "Cloning and Characterization of a binding subunit of the interleukin 13 receptor that is also a component of the interleukin 4 receptor" p.497-501 See patent family annex. Further documents are listed in the continuation of Box C. Special cargories of cited documents: document deflaing the general state of the art which is not considered to be of particular relevance earlier document but published on or wher the international filling date document which may throw doubts on priority cisim(s) or which in cited to establish the publication date of another cistation or other special reason (as specified) document referring to an oral disclosure, use, exhibition or other later document published after the interactional filing date or priority date and not in conflict with the application but cited to und the principle or theory underlying the invention document of particular relevance; the claimed inver considered sovel or cannot be considered to involve an inventive step then the document is taken alone document of particular relevance, the chimed invention cannot be nt referring to an oral disclosure, use, exhibition or other considered to involve an inventive step when the document is combined with one or more other such documents, such combined °o° document published prior to the interactional filing date but inter then the priority date chained being obvious to a person skilled in the art document member of the same patent family Date of the actual completion of the international search Date of mailing of the international search report 24 September, 1999 (24. 09. 99) 5 October, 1999 (05. 10. 99) Name and mailing address of the ISA/ Authorized officer Japanese Patent Office Telephone No.

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